

Original Article

DOCKING AND CYTOTOXICITY STUDIES OF 2-VINYLCROMONE DERIVATIVES ON HUMAN BREAST CANCER CELL LINES

SWATI KAUSHIK#, MEGHA RIKHI#, SEEMA BHATNAGAR*

Novel Molecule Synthesis Laboratory, Amity Institute of Biotechnology, Amity University, Noida 201303, Uttar Pradesh, India
Email: sbhatnagar1@amity.edu

Received: 16 Aug 2015 Revised and Accepted: 27 Oct 2015

ABSTRACT

Objective: Estrogen receptor (ER) is over-expressed in 70% of breast cancers. The ER has two isoforms, ER α and ER β . The ER ligand binding domain (LBD) has been the target for hormone-responsive breast cancer. Due to tissue-specific effects currently available drugs for hormone positive breast cancer presents serious limitation. The dynamic and plastic nature of ER LBD plays a crucial role in ligand design that discriminates between the ER subtypes. Agents that selectively target ER isoform are a formidable challenge to researchers. The chromone scaffold is a privileged scaffold for exploration of anticancer agents. The objective of the present study was to evaluate the anticancer activity of a small library of 2-vinylchromones in human breast cancer cell lines MCF-7 and MDA-MB-231.

Methods: The compounds were synthesized by the reported procedures. Docking studies of the substituted 2-vinylchromone was performed using GLIDE tool in Maestro 8.0. The compounds were evaluated for anticancer activity against MCF-7 (ER α positive), MDA-MB-231 (ER β positive) and MRC-5 (ER α , β negative) cell lines using MTT assay.

Results: The *in silico* studies indicated that substituted 2-vinylchromones, **1(a-c)** and **2(a-b)** exhibited comparable docking score at LBD of ER α and ER β . However, the binding affinity of the compounds for the allosteric binding site in ER β was negligible. The dose-dependent studies using MTT assay depicted that compounds **1(a-c)** and **2(a-b)** exhibited anticancer activity in ER α positive cell line MCF-7 as compared to ER β positive cell line **MDA MB 231**. The most potent anticancer activity was observed for compound **2b** against MCF-7 cells with IC₅₀ value of 15.625 μ g/ml.

Conclusion: The present investigation indicated that 2-vinylchromone derivatives exhibited ER isoform selectivity and the presence of bulky group in 2-vinylchromones resulted in significantly higher cytotoxicity in ER α positive cell lines as compared to the ER β positive cell line.

Keywords: Estrogen receptor, 2-vinylchromone, Docking, Anticancer, MCF-7, MDA-MB-231.

INTRODUCTION

Chromones are an important class of oxygen-containing heterocyclic compounds. Synthetic and natural chromone derivatives possess important biological activities such as antitumor [1], antihepatotoxic [2], antioxidant [3], anti-inflammatory [4], antiallergic [5], estrogenic [6] and antibacterial [7] activities. The anticancer activity of chromones has also been related to its structural similarity with flavonoids.

Several compounds such as quercetin, genistein, daidzein, apigenin, biochanin A, kaempferol and naringenin that have chromone scaffold are known to bind to ER isoforms. Genistein, apigenin and kaempferol have a higher binding affinity 20 to 30-fold more for ER β than ER α as indicated by a solid-phase binding assay [8-9].

The above literature precedence prompted us to investigate the docking and cytotoxicity of 2-vinylchromones on human breast cancer cell lines. Previous work in our own group had led to the synthesis of novel chromone derivatives such as 3,4-dihydro-2H-1-benzopyran-3,4 dione **1(a-c)** using NaBH₄ (unpublished results) and in another study E-3-aminochromen-4-one **2(a-b)** were synthesized (unpublished results). Herein, we report the *in silico* studies of chromone derivatives along with an assessment of cytotoxic activity of the compounds on MCF-7, MDA-MB-231 and MRC-5 cells.

MATERIALS AND METHODS

Determination of anticancer activity

MTT assay

5x10³ cells/well were seeded in 96-multiwell flat bottom microtitre plate and cultured overnight in a humidified atmosphere of 5% CO₂ at 37 °C. The test compounds were serially diluted to the wells. Different concentrations ranging from 125 μ g/ml to 0.98 μ g/ml were analyzed. The cells were incubated for 24 h with test compounds

with 5% CO₂ at 37 °C. After 24 h, 10 μ l of MTT (5 mg/ml in 1X PBS) was added to each well. The cells were incubated for 3 h and the centrifuged at 1500 rpm for 10 min. The media were discarded and 150 μ l of DMSO was added to each well. Optical density was measured using a microplate reader at 570 nm with reference optical density at 630 nm. IC₅₀ values were calculated as the concentrations that show 50% inhibition of proliferation on the tested cell line.

Docking methodology

The compounds were modelled using the BUILD application of Maestro 8.0. The geometry was optimized by molecular mechanics using IMPACT in a dynamic environment using the standard TIP4P water model. The energy minimization was done using Optimized Potentials for Liquid Simulations 2005 (OPLS 2005) force field. Energy minimization was done using Polak-Ribier conjugate gradient and Truncated Newton conjugate gradient algorithms. The convergence threshold used was rms gradient of 0.01.

Conformational models of the ligands were generated. Docking of the ligands was carried out using an extra precision (XP) method called GLIDE (Grid-based Ligand Docking with Energetics) for flexible ligand docking. The ligands were prepared for docking using LIGPREP from Schrodinger's molecular modeling software [10]. The PDB entries chosen for the above studies were 2QTU, 2FSZ and 3ERT. The ligand interaction map was used to study the interactions of novel molecules with the residues of ER.

Statistical analysis

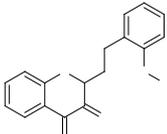
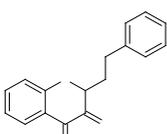
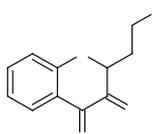
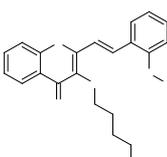
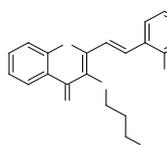
The experimental data obtained for each set of the experiment were expressed as the mean of change \pm SEM and analyzed by one-way and two-way ANOVA followed by a bonferroni post-test. Level of significance was set to P \leq 0.05. All the statistical calculations were performed using the evaluation version of Graph Pad Prism 5.1 statistical software.

RESULTS

The synthesized compounds were evaluated for their binding affinity towards ER α and ER β by docking studies. The PDB entry chosen for the docking studies were 2QTU [11], 2FSZ [12] and 3ERT [13]. The PDB entry 3ERT is a complex of ER α with hydroxy-tamoxifen (HT). The selection of 2QTU entry was based on structural similarity of the synthesized ligands with benzopyranones. Native ER β pdb entry is not available therefore 2FSZ was selected which is a complex of HT with ER β . The compounds were modelled using the BUILD application of Maestro 9.6. The energy minimization was performed using optimized potentials for liquid simulations 2005 (OPLS 2005) force fields. In case of ER β isoform, the docking studies were performed at the cognate site (2FSZ-101) as well as the allosteric binding site (2FSZ-103) of HT. The docking score of compounds **1(a-c)** and **2(a-b)** are indicated in table 1.

The docking studies indicated that **1(a-c)** and **2(a-b)** exhibited comparable docking score at LBD of ER α and ER β . However, the binding affinity of the compounds for the allosteric binding site in ER β [12] was negligible. Amongst the analogues synthesized, best docking score for ER α was exhibited by compound 3-(butylamino)-2-[(E)-2-(2-methoxyphenyl) ethenyl]-4H-chromen-4-one (**2b**) containing aminoalkyl group and 2-[2-(2-methoxy phenyl)ethyl]-3,4-dihydro-2H-1-benzopyran-3,4 dione (**1b**) having a keto group at C-3 position. A comparison of the ligand interaction map represented in fig. 1 indicated that majority of the poses obtained for **2b** share the following common residues: Glu 353, Arg 394, Trp 383, Ala 350, Met 343, Leu 391, Ile 424, Leu 525, Leu 346, Leu 387 as compared with HT. Compound **2b** also depicted other significant interactions with the following residues: Asp 351, Leu 349, Leu 354, Leu 536, Met 528, Thr 347, His 524, Leu 428, Met 388, Leu 384, Phe 404, Ala 350.

Table 1: Docking score and glide energy of the compounds **1(a-c)** and **2(a-b)** against ER α and ER β

S. No.	Compound	Structure	3ERT		2QTU		2FSZ (101)		2FSZ (103)	
			Docking Score	Glide energy of the model (Kcal/mol)	Docking Score	Glide energy of the model (Kcal/mol)	Docking Score	Glide energy of the model (Kcal/mol)	Docking Score	Glide energy of the model (Kcal/mol)
1	1a		-5.19	-36.75	-7.8	-40.29	-6.57	-34.87	-3.42	-23.76
2	1b		-7.1	-27.4	-7.78	-38.64	-6.7	-36.56	-4.25	-29.98
3	1c		-6.14	-26.81	-6.03	-30.17	-5.23	-27.11	-3.42	-20.63
4	2a		-6.4	-34.35	-3.09	-21.86	-6.83	-43.35	-4.61	-30.49
5	2b		-8.27	-55.59	-3.87	-24.97	-5.84	-39.08	-3.39	-26.98

3ERT is a complex of ER α with HT. 2QTU pdb entry is a complex of benzopyranones with ER β . 2FSZ is a complex of HT with ER β where docking studies performed at the cognate site of HT was labeled as 2FSZ-101 and the allosteric binding site was labeled as 2FSZ-103.

The glide energy of the compound **1b** and **2b** with ER α was found to be -27.40Kcal/mol and -55.59Kcal/mol with -7.01 and -8.27 docking score. Fig. 1 (A and B) illustrates the interaction of compound **1b** and **2b** with amino acid residues of ER α (PDB ID 3ERT).

The dose-dependent studies using MTT assay depicted that compounds **1(a-c)** and **2(a-b)** exhibited anticancer activity in ER α positive cell line MCF-7 as compared to ER β positive cell

line MDA-MB-231. The compound **1b**, **2a** and **2b** exhibited anticancer activity in ER α positive cell line. Amongst them compound **2b** was most potent. Remarkable differences in the IC₅₀ value were seen for **1b**, **2a** and **2b** in MCF-7 and MDA-MB-231. The IC₅₀ value for **1b**, **2a**, **2b** were 62.5 μ g/ml, 31.25 μ g/ml and 15.625 μ g/ml, respectively in MCF-7 cell line. All the compounds exhibited an IC₅₀ value of more than 125 μ g/ml in MDA-MB-231 cell line as indicated in table 2.

In an attempt to corroborate the results of docking studies for compounds **1(a-c)** and **2(a-b)** MTT assay were performed in ER α and ER β positive cell lines. The cell lines chosen for the studies were MCF-7 (ER α positive), MDA-MB-231 (ER β positive) and MRC-5 (ER α , β negative). The results of these studies are depicted in fig. 4.

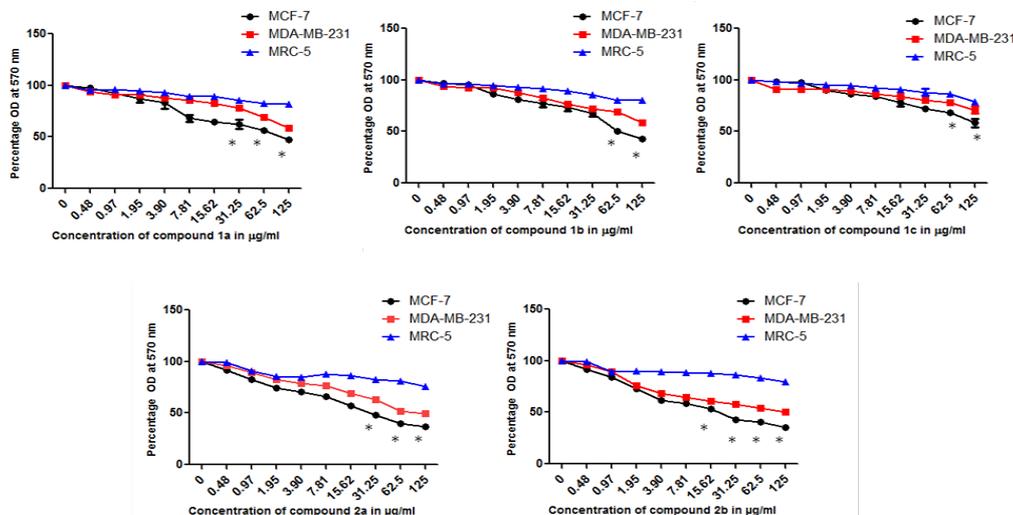


Fig. 4: Comparative graph for 1(a-c) and 2(a-b) compounds with % change in optical density in respective cell lines plotted against different concentrations of compounds. The data are mean \pm SEM from 2 samples for each group and analyzed by two-way ANOVA followed by Bonferroni Post-test where * $P < 0.01$ significant from control

DISCUSSION

The LBD of ER α and ER β has 60% homology [14]. The dynamic and plastic nature of ER LBD plays a crucial role in ligand design that discriminates between the ER subtypes.

The interior of the ligand binding pocket of ER α and ER β have been reported to have 22-24 residues which are identical and involved in interactions with ligands [15]. The two pockets are however different in size and flexibility and this aspect is crucial for the development of selective subtype agents [16].

Apart from this the discovery of allosteric binding pocket in the coactivator groove of ER β further provides avenues for identification of agents known as coactivator binding inhibitors [12, 17]. In the present exploratory work we have attempted to identify ER isoform-selective compounds from a small library of 2-vinylchromones. For this purpose docking studies have been carried at both ER α and ER β LBD and allosteric binding pocket of ER β . The findings of docking studies have been correlated with MTT assay on ER α , ER β and ER α , β negative cell lines.

The *in silico* studies revealed that the synthesized compounds preferentially binds to the ligand binding domain of ER α and ER β with a comparable docking score except for **2b** which exhibits preferential docking to ER α . It was also found that these compounds exhibited poor binding to the allosteric binding site of ER β . Highest docking score amongst the series was also observed for compound **2b**. Prominent interacting residues including Glu 353, Arg 394 and Asp351 as compared to HT clearly indicate high-affinity binding which is further corroborated with a high docking score of **2b**. A comparison of docking residues of ER β , on the other hand, did not reveal an interaction with these residues.

Further, the evaluation of the cytotoxic activity indicated that these novel compounds have exhibited significant cytotoxicity in ER α positive cell line instead of ER β positive cell line. Studies on ligands such as genistein have clearly established that although it exhibits similar binding mode when complexed to ER α and ER β [18] it exhibits 40-fold selectivity for ER β [19]. Similarly, electron density maps of HT complexed to ER β indicate one of the HT molecules is located in the cognate ligand binding pocket with a confirmation indistinguishable from ER α /HT structure [20]. However; a comparison of IC₅₀ values of

HT in ER α and ER β indicates ER α selectivity [21]. These observations suggest the involvement of different residues of ER α and ER β LBD that may contribute towards isoform selectivity.

In our studies, it was observed that in all cases the chromones with methoxy substitution **1b**, **2a** and **2b** exhibited the highest cytotoxicity. The presence of a bulky aminoalkyl group at C-3 of the chromone ring as compared to the keto group as in the case of **2a** and **2b** respectively led to significantly higher cytotoxicity. Further; structure-activity relationship studies will help to correlate contribution of long chain aminoalkyl substitution to cytotoxicity.

CONCLUSION

2-vinylchromone derivatives exhibited higher cytotoxicity in ER α positive cell lines as compared to the ER β positive cell line. Our results also indicated that these derivatives exhibited ER isoform selectivity and presence of bulky group as in the case of **2a** and **2b** resulted in a significant increase in cytotoxic activity.

ACKNOWLEDGEMENT

The authors gratefully acknowledge Amity University for infrastructure and facilities. Author, MR thanks financial assistance through Amity Science, Technology and Innovation Foundation (ASTIF) fellowship provided by Amity University. Author, SK thanks Indian Council of Medical Research (ICMR) for financial support through Senior Research Fellowship (SRF).

CONFLICT OF INTERESTS

Declared None

REFERENCES

- Marten's S, Mithofer A. Flavones and flavone synthases. *Phytochemistry* 2005;66:2399-407.
- Fernandes E, Carvalho M, Carvalho F, Silva AMS, Santos CMM, Pinto DCGA, et al. Hepatoprotective activity of polyhydroxylated 2-styrylchromones against tert-butylhydroperoxide induced toxicity in freshly isolated rat hepatocytes. *Arch Toxicol* 2003;77:500-3.
- Kuroda M, Uchida S, Watanabe K, Mimaki K. Chromones from the tubers of *eranthis ciliica* and their antioxidant activity. *Phytochemistry* 2009;70:288-93.

4. Gabor M. Anti-inflammatory and anti-allergic properties of flavonoids. *Prog Clin Biol Res* 1986;213:471-80.
5. Doria G, Romeo C, Forgiome A, Sberze P, Tibolla N, Corno ML, *et al.* Antiallergic agents. III. Substituted trans-2-ethenyl-4-oxo-4H-1-benzopyran-6-carboxylic acids. *Eur J Med Chem* 1979;14:347-51.
6. Maggioline M, Bonofiglio D, Marisco S, Panno ML, Cenni B, Picard D, *et al.* Estrogen receptor alpha mediates the proliferative but not the cytotoxic dose-dependent effects of two major phytoestrogens on human breast cancer cells. *Mol Pharmacol* 2001;60:595-602.
7. Grindlay D, Reynolds T. The aloe vera phenomenon: a review of the properties and modern uses of the leaf parenchyma gel. *J Ethnopharmacol* 1986;16:117-51.
8. Kuiper GG, Carlsson B, Grandien K, Enmark E, Haggblad J, Nilsson S, *et al.* Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptors alpha and beta. *Endocrinology* 1997;138:863-70.
9. Kuiper GG, Lemmen JG, Carlsson B, Corton JC, Safe SH, Van der Saag PT, *et al.* Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology* 1998;139:4252-63.
10. Friesner RA, Banks JL, Murphy RB, Halgren TA, Klicic JJ, Mainz DT, *et al.* Glide: a new approach for rapid, Accurate docking and scoring. 1. Method and assessment of docking accuracy. *J Med Chem* 2004;47:1739-49.
11. Richardson TI, Dodge JA, Wang Y, Durbin JD, Krishnan V, Norman BH. Benzopyrans as selective estrogen receptor beta agonists (SERBAs). Part 5: Combined A-and C-ring structure-activity relationship studies. *Bioorg Med Chem Lett* 2007;17:5563-6.
12. Wang Y, Chirgadze NY, Briggs SL, Khan S, Jensen EV, Burris TP. A second binding site for hydroxytamoxifen within the coactivator-binding groove of estrogen receptor beta. *Proc Natl Acad Sci* 2006;103:9908-11.
13. Shiau AK, Barstad D, Loria PM, Cheng L, Kushner PJ, Agard DA, *et al.* The structural basis of estrogen receptor/coactivator recognition and the antagonism of this interaction by tamoxifen. *Cell* 1998;95:927-37.
14. Petit FG, Valotaire Y, Pakdel F. The analysis of chimeric human/rainbow trout estrogen receptors reveals amino acid residues outside of P-and D-boxes important for the transactivation function. *Nucleic Acids Res* 2000;28:2634-42.
15. Pike AC, Brzozowski AM, Hubbard RE, Bonn T, Thorsell AG, Engstrom O, *et al.* Structure of the ligand-binding domain of estrogen receptor beta in the presence of a partial agonist and a full antagonist. *EMBO J* 1999;18:4608-18.
16. Katzenellenbogen JA, Muthyala R, Katzenellenbogen BS. The nature of the ligand-binding pocket of estrogen receptor alpha and beta: the search for subtype-selective ligands and implications for the prediction of estrogenic activity. *Pure Appl Chem* 2003;75:2397-403.
17. Sun A, Moore TW, Gunther JR, Kim MS, Rhoden E, Du Y, *et al.* Discovering small-molecule estrogen receptor α /coactivator binding inhibitors: high-throughput screening, ligand development, and models for enhanced potency. *ChemMedChem* 2011;6:654-66.
18. Manas ES, Xu ZB, Unwalla RJ, Somers WS. Understanding the selectivity of genistein for human estrogen receptor-beta using X-ray crystallography and computational methods. *Structure* 2004;12:2197-207.
19. Poluzzi E, Piccinni C, Raschi E, Rampa A, Recanatini M, Ponti FD. Phytoestrogens in postmenopause: the state of the art from a chemical, pharmacological and regulatory perspective. *Curr Med Chem* 2014;21:417-36.
20. Kojetin DJ, Burris TP, Jensen EV, Khan SA. Implications of the binding of tamoxifen to the coactivator recognition site of the estrogen receptor. *Endocr-Relat Cancer* 2008;15:851-70.
21. Weatherman RV, Clegg NJ, Scanlan TS. Differential SERM activation of the estrogen receptors (ER α and ER β) at AP-1 sites. *Chem Biol* 2001;8:427-36.