

Original Article

PICEATANNOL AMELIORATES CISPLATIN-INDUCED HISTOLOGICAL AND BIOCHEMICAL ALTERATIONS IN RATS KIDNEY

SARA A. WAHDAN^a, SAMAR S. AZAB^a, DOAA A. ELSHERBĪNY^a, EBTEHAL EL-DEMERDASH^a

^aPharmacology and Toxicology Department, Faculty of Pharmacy, Ain shams University, Cairo, Egypt
Email: ebtehal_dm@yahoo.com

Received: 04 Oct 2016 Revised and Accepted: 31 Aug 2017

ABSTRACT

Objective: The present study was designed to investigate the effect of different doses of piceatannol (PIC) on cisplatin-induced biochemical and histological alterations in rat kidney.

Methods: Male Wistar rats received a single intraperitoneal (i. p.) injection of cisplatin (7 mg/kg). PIC was given in different daily doses (5, 10 and 20 mg/kg) i. p., for seven days, starting two days before cisplatin injection. Nephrotoxicity was evaluated by means of measurement of blood urea nitrogen (BUN), serum creatinine and histopathological examination of the kidney. We also investigated the antioxidant effect of the most effective dose of PIC by measuring reduced glutathione (GSH) and lipid peroxides levels. Moreover, the ability of PIC to affect nuclear factor kappa B (NF-κB) expression was determined by immunohistochemical analysis.

Results: A single dose of cisplatin (7 mg/kg) significantly increased BUN and creatinine levels, as well as relative kidney weight, compared to the control group. In addition, significant histopathological alterations including tubular necrosis, hemorrhage and casts formation were observed. PIC was given in different doses (5, 10 and 20 mg/kg) for 7 d, starting 2 d before cisplatin injection. PIC dose 10 mg/kg was the most effective in preventing these alterations. PIC significantly increased GSH level and decreased lipids peroxidation compared to cisplatin group. Moreover, PIC significantly mitigated cisplatin-induced expression of NF-κB.

Conclusion: PIC has the potential to ameliorate cisplatin-induced renal injury.

Keywords: Cisplatin, Piceatannol, Nephrotoxicity, Antioxidants, NF-κB

© 2017 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>)
DOI: <http://dx.doi.org/10.22159/ijpps.2017v9i10.15518>

INTRODUCTION

Cisplatin is an effective and widely used chemotherapeutic agent [1]. Nephrotoxicity represents one of its major and clinically significant side effects, limiting its clinical use [2]. The high prevalence of cisplatin-induced nephrotoxicity is well documented, occurring in about one-third of patients receiving cisplatin treatment [3]. Cisplatin-induced renal toxicity is dose-dependent, limiting the possibility of increasing dosages, which affects treatment effectiveness [4]. It has a direct toxic effect on renal tubules, vasculature and glomeruli. Multiple cellular mechanisms were found to be involved in cisplatin-induced renal damage, including oxidative stress, inflammatory response and activation of apoptotic pathways [5]. Clinically, renal toxicity begins with 20% to 40% decline in glomerular filtration, followed by increased serum levels of creatinine and blood urea nitrogen (BUN), hypomagnesemia and hypokalemia [1].

There is no specific treatment for the cisplatin-induced renal injury. The most important supportive measures may include adequate hydration, use of diuretics and electrolytes replacement [6]. Therefore, considerable efforts have been focused on exploring new compounds serving as protective treatments against cisplatin-induced nephrotoxicity [7, 8].

Piceatannol (3,4',3',5'-trastrihydroxystilbene) (PIC) is a natural analogue and a metabolite of resveratrol [9]. It was first isolated from the seeds of *Euphorbia lagascae* [10]. The most important sources of PIC in the diet are grapes, passion fruit, white tea, and Japanese knotweed [11]. PIC is an important naturally occurring polyphenolic stilbene, produced by plants in response to fungal infection, mechanical damage and ultra-violet irradiation. It can be consumed on a daily basis with no harmful effects on human body [12]. PIC is known as a potent scavenger of free radicals due to hydroxyl groups in its stilbene rings [13]. The anti-proliferative, anti-inflammatory and antioxidant properties of PIC were proved in

multiple *in vivo* and *in vitro* studies [14-17]. One study showed that PIC is a potent enhancer of cisplatin sensitivity in ovarian cancer, potentially, through the modulation of several major determinants of chemoresistance [18]. However, its role as a protective agent against cisplatin-induced nephrotoxicity has not been investigated. Therefore, the present study was performed to investigate the potential protective effect of different doses of PIC against cisplatin-induced nephrotoxicity in rats.

MATERIALS AND METHODS

Animals

Male Wistar rats weighing 200–250 g were purchased from the National Institute of Research, (Cairo, Egypt). Rats were kept in an air-conditioned atmosphere, at a temperature of 25 °C with alternatively 12 h light and dark cycles and kept on a standard diet *ad libitum* and water. Standard diet pellets (El-Nasr, Abu Zaabal, Egypt) containing not less than 20% protein, 5% fiber, 3.5% fat, 6.5% ash and a vitamin mixture. Few days before the experiment, animals were acclimatized to laboratory conditions. The study was approved by the ethical committee of Faculty of Pharmacy, Ain Shams University (Cairo, Egypt) (REC-ASU PhD No. 5).

Drugs and chemicals

Cisplatin was obtained from Mylan (Saint-Priest, France), dissolved in saline at a concentration 1 mg/ml and injected i. p. as a single dose of 7 mg/kg. PIC (98% pure) was purchased from Alexis Biochemicals (San Diego, CA, USA) and dissolved in dimethyl sulfoxide (DMSO) and normal saline (0.9%) in a ratio (2:3). The protective effect of different doses of PIC (5, 10 and 20 mg/kg) against cisplatin-induced nephrotoxicity was screened by the assessment of nephrotoxicity markers. Serum creatinine and BUN were estimated using available commercial kits purchased from Endomedics Co. (Cairo, Egypt). GSH and lipid peroxides levels were determined using commercial kits obtained from Biodiagnostics Co. (Giza, Egypt). Rabbit polyclonal

anti NF- κ B p65 antibody was purchased from Thermo Fisher Scientific (USA) (Cat No. RB-9034-P). Bovine serum albumin, diaminobenzidine, tris base and DMSO were purchased from Sigma Chemical Co. (St. Louis, MO, USA). Ethanol, methanol, formaldehyde, dipotassium hydrogen phosphate (K_2HPO_4) and potassium dihydrogen phosphate (KH_2PO_4) were purchased from El Nasr Chemical Co. (Cairo, Egypt). All other chemicals were of the highest purity grade commercially available.

Experimental design

Screening the effective protective dose of PIC against cisplatin-induced nephrotoxicity

A dose response screening study was conducted, in which animals were randomly divided into six groups (ten animals per group) and treated as follows; The first group was considered as control group and received the vehicle (DMSO and isotonic saline) (2:3), 0.2 ml/200 gm body weight (i. p.), once daily for 7 consecutive days and a single injection of isotonic saline (0.2 ml/200 gm body weight, i. p.) on the third day of the experiment. The second group was given the vehicle i. p., once daily for 7 consecutive days and a single injection of cisplatin (7 mg/kg, i. p.) on the third day of the experiment to induce nephrotoxicity. The third, fourth and the fifth groups received PIC at a dose of 5, 10 and 20 mg/kg, respectively, i. p., once daily for 7 consecutive days and a single dose of cisplatin 7 mg/kg i. p. on the third day of the experiment. PIC was given 2 h prior to cisplatin injection [16]. The sixth group received PIC at a dose 20 mg/kg i. p., once daily for 7 consecutive days.

At the end of the experiment, the number of dead animals and the change in animals' weight were recorded. Then, rats were anesthetized and blood samples were collected from the retro-orbital plexus and allowed to clot. Serum was separated by centrifugation at 3000 rpm for 10 min and was stored at -80 °C until assayed for the biochemical parameters. Then rats were sacrificed and kidney tissues were dissected out and washed with ice-cold saline. Additionally, specimens from kidneys of each group were fixed in 10 % formalin for histopathological examination.

Studying the possible mechanisms underlying the protective effect of PIC against cisplatin-induced nephrotoxicity

In order to determine the potential mechanism underlying the nephroprotective effect of PIC, another study was conducted using its most effective dose in which animals were divided randomly into 4 groups (ten animals per group) and treated as follows; The first group was considered as control group and given the vehicle (DMSO and isotonic saline) (2:3) i. p., once daily for 7 consecutive days and a single injection of isotonic saline i. p. on the third day of the experiment. The second group was given the vehicle i. p., once daily for 7 consecutive days and a single injection of cisplatin (7 mg/kg, i. p.) on the third day of the experiment to induce nephrotoxicity. The third, group received PIC at a dose 10 mg/kg, i. p., once daily for 7 consecutive days and a single dose of cisplatin 7 mg/kg i. p. on the third day of the experiment. The fourth group received PIC at a dose 10 mg/kg i. p., once daily for 7 consecutive days.

At the end of the experiment, kidney tissues were dissected and washed with ice-cold saline. Portions of the kidney tissues were homogenized in 0.1 M phosphate buffer (pH 7.4) producing 10% homogenate. The homogenates were centrifuged at 4000 rpm at 4 °C for 15 min then aliquots of supernatants were separated and used for biochemical analyses. Additionally, specimens from kidneys of each group were fixed in 10 % formalin for immunohistochemical analysis.

Assessment of nephrotoxicity markers

Serum creatinine and BUN were estimated using available commercial kits (Endomedics Co. Cairo, Egypt). Briefly, creatinine was determined according to Jaffe-kinetic method which depends on the measurement of the rate of formation of a colored complex between creatinine and alkaline picrate [19]. BUN determination depends on the formation of a green colored compound due to the reaction between urea and Na nitroprusside in the presence of urease enzyme [20]. The intensity of this color is proportional to BUN. Spectrophotometric analyses were performed using UV-visible

spectrophotometer (UV-1601) (Shimadzu Co, Japan). Relative kidney weight was calculated according to the formula: (kidney weight/total body weight) \times 100.

Histopathological examination

For light microscopy, kidney specimens were fixed in 10% formalin for 24 h and then washed with tap water. Serial dilutions of alcohol (methyl, ethyl and absolute ethyl alcohol) were used for dehydration. Specimens were cleared in xylene embedded in paraffin at 56 °C in hot air oven for 24h. Paraffin tissue blocks were prepared for sectioning at 4 microns thickness by sledge microtome. The obtained tissue sections were collected on glass slides and deparaffinized. Finally, sections were stained with hematoxylin and eosin (H and E) and examined under the light microscope (Olympus BX-50 Olympus Corporation, Tokyo, Japan). [21].

Assessment of oxidative stress markers

Assessments of reduced glutathione (GSH) and lipid peroxides levels were performed using commercial kits (Biodiagnostics Co. Giza, Egypt). Briefly, GSH assessment was based on the reduction of 2-nitrobenzoic acid by GSH to produce a yellow compound. The absorbance of the yellow colored product was determined at 405 nm. Lipid peroxidation was estimated spectrophotometrically by measuring malondialdehyde (MDA) level. Thiobarbituric acid (TBA) reacts with MDA in acidic medium at temperature 95 °C for 30 min to form a thiobarbituric acid reactive product. The absorbance of the pink colored product was determined at 535 nm. Spectrophotometric analyses were performed using UV-visible spectrophotometer (UV-1601) (Shimadzu Co, Japan).

Immunohistochemical detection of NF κ B (p65)

Paraffin-embedded tissue sections, of 3 μ m thickness, were rehydrated first in xylene and then in graded ethanol solutions. The slides were then blocked with 5% bovine serum albumin in tris-buffered saline for 2 h. The sections were then immunostained with the rabbit polyclonal anti NF- κ B p65 primary antibody (Thermo Fisher Scientific, Cat No. RB-9034-P), at a concentration of 1 μ g/ml containing 5% bovine serum albumin in tris-buffered saline and incubated overnight at 41 °C. After washing the slides with tris-buffered saline, the sections were incubated with goat anti-rabbit secondary antibody. Sections were then washed with tris buffered saline and incubated for 5–10 min in a solution of 0.02% diaminobenzidine containing 0.01% H_2O_2 . Counter staining was performed using hematoxylin stain. Slides were examined with the light microscope (Olympus BX-50 Olympus Corporation, Tokyo, Japan). Immunohistochemical quantification was carried out using image analysis software (Image J, 1.46a, NIH, USA).

Statistical analysis

Data were presented as mean \pm SD Statistical analysis was performed using one-way analysis of variance (ANOVA), followed by Tukey-Kramer as a post Hoc test. Two-tailed Student's t-test was used for statistical comparisons between two groups. The 0.05 level of probability was used as the criterion for significance. All statistical analyses were performed using Instat software package (version 3). Graphs were sketched using GraphPad Prism software (version 5).

RESULTS

Mortality, body weight and relative kidney weight

The results of nephrotoxicity markers including mortality, total body weight and relative kidney weight were shown in table 1. In cisplatin group, three animals died and a significant decrease in the body weight of rats by 17%, as compared to their initial weight associated with a significant increase in relative kidney weight by 32%, as compared to the control group, were observed. Two animals died in the group treated concomitantly with PIC (5 mg/kg). No animal death was observed in rats co-treated with 10 and 20 mg/kg PIC. Other treated groups did not show any significant change in their body weight, as compared to their initial weight or in relative kidney weight, as compared to the control group. Concomitant treatment with PIC (5, 10 and 20 mg/kg) caused a significant reduction in relative kidney weight by 22%, 30% and 30%, respectively, as compared to cisplatin-treated group.

Table 1: Effects of different doses of piceatannol (PIC) on body weight and relative kidney weight in cisplatin-treated rats

| Groups | No. of dead rats | Body weight (gms) | | Relative kidney weight (%) |
|--------------------------|------------------|-------------------|-----------------|----------------------------|
| | | Before treatment | After treatment | |
| Control | 0/10 | 221±40.14 | 242±38.3 | 0.68±0.07 |
| Cisplatin | 3/10 | 214.9±45.8 | 178.9±16.3* | 1±0.178 ^a |
| Cisplatin+PIC (5 mg/kg) | 2/10 | 240.7±45.9 | 202.6±25.7 | 0.78±0.16 ^b |
| Cisplatin+PIC (10 mg/kg) | 0/10 | 260±57.9 | 215±32.8 | 0.7±0.05 ^b |
| Cisplatin+PIC (20 mg/kg) | 0/10 | 230±34.6 | 210.8±43.5 | 0.7±0.04 ^b |
| PIC (20 mg/kg) | 0/10 | 210.5±52.7 | 220±39.7 | 0.64±0.02 ^b |

Data were presented as means±SD (n=10), a: significantly different from the control group; b: significantly different from cisplatin group. P<0.05 using ANOVA followed by Tukey-Kramer as a post-hoc test. *Significantly different from pre-treated group, P<0.05 using student paired t-test, PIC: Piceatannol.

Serum creatinine and BUN

Cisplatin treatment caused a significant increase in serum creatinine and BUN by 564% and 764%, respectively, compared to the control group. Concomitant treatment with PIC (5, 10 and 20 mg/kg) significantly ameliorated nephrotoxicity markers, indicated by a significant reduction in serum creatinine levels by 27, 55 and 53%, respectively, as compared to cisplatin treated group. Similarly, treatment with PIC (5, 10 and 20 mg/kg) significantly reduced BUN

levels by 33, 58 and 50%, respectively, compared to cisplatin group. In addition, serum creatinine and BUN in animals treated with PIC (10 and 20 mg/kg) were significantly lower than those in the group treated with PIC (5 mg/kg). On the other hand, there was no significant difference between the two groups treated with PIC (10 mg/kg) and PIC (20 mg/kg) in all nephrotoxicity markers. Treatment of rats with PIC alone did not show any significant change in all nephrotoxicity parameters, as compared to the control group (fig. 1).

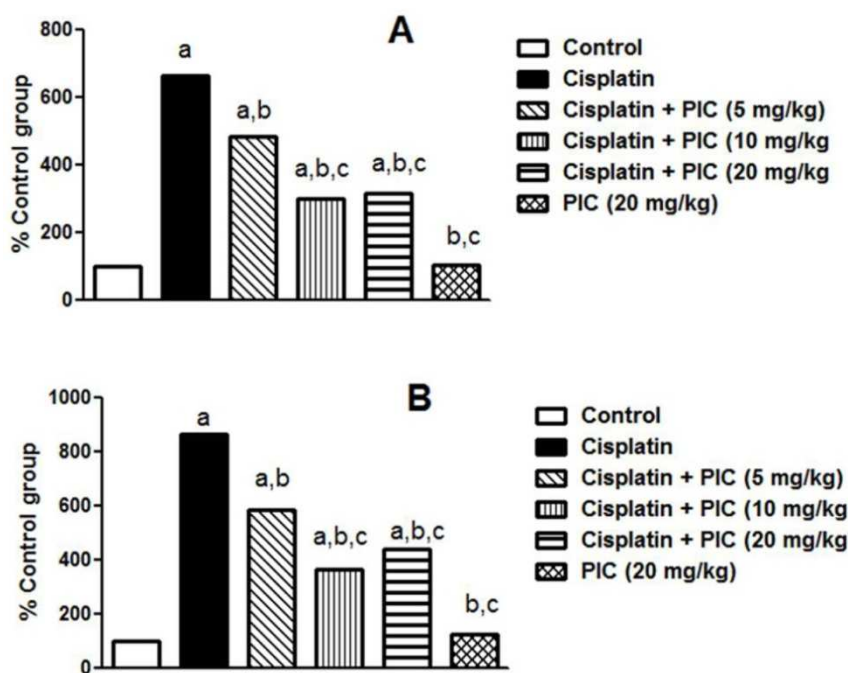


Fig. 1: Effects of different doses of piceatannol (PIC) on (A) serum creatinine and (B) BUN in cisplatin-treated rats, Data were presented as % of control group, a: significantly different from the control group; b: significantly different from cisplatin group; c: significantly different from cisplatin and PIC (5 mg/kg) treated group. P<0.05 using ANOVA followed by Tukey-Kramer as a post-hoc test

Histopathological examination

Histopathological alterations in kidney specimens from different treatment groups were shown in fig. 2. Kidney specimens from the control rats (fig. 2A) and rats treated with PIC (20 mg/kg) alone (fig. 2B) showed a normal histological structure of the glomeruli and renal tubules. Treatment of rats with cisplatin caused severe congestion in the cortical blood vessels, as well as in the glomerular tufts and was associated with periglomerular inflammatory cells aggregation (fig. 2C). Degeneration in the tubular epithelial cells and eosinophilic casts formation in the tubular lumen were also observed in the cortical portion (fig. 2D). Kidney specimens from

rats treated with PIC (5 mg/kg) concomitantly with cisplatin showed mild reduction in cisplatin-induced pathological alterations. Focal inflammatory cells, tubular necrosis, homogenous eosinophilic casts formation and cystic dilatation were observed (fig. 2E). Concomitant treatment of rats with PIC at higher doses (10 and 20 mg/kg) ameliorated the pathological changes induced by cisplatin and restored the normal histological structure of the glomeruli and tubules (fig. 2F and G). Only mild focal hemorrhage in the cortex was detected in the kidney specimens from the group pre-and co-treated with PIC (10 mg/kg) (fig. 2F). Various aspects of histopathological alterations in kidney specimens which were graded, as shown in table 2.

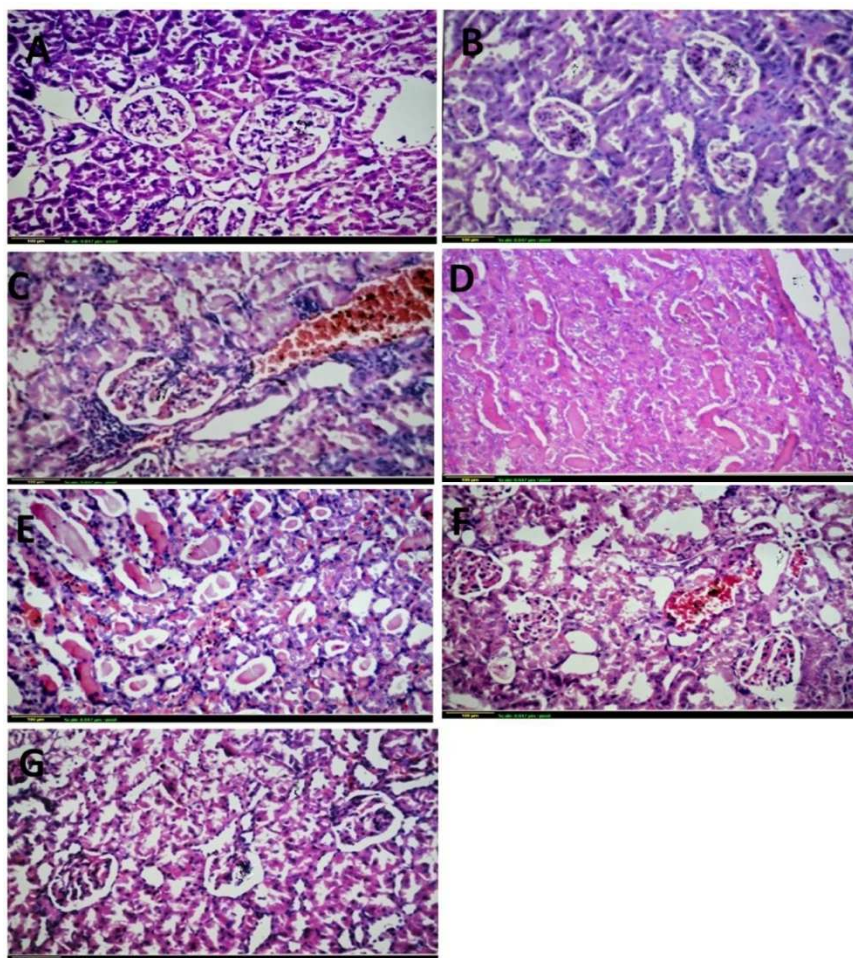


Fig. 2: Effects of different doses of piceatannol (PIC) on cisplatin-induced histological alterations of the kidney tissue, Representative photomicrographs of kidney sections stained with hematoxylin-eosin stain (magnification ×40) (A) control group, (B) PIC alone (20 mg/kg) treated group, (C and D) Cisplatin treated group, (E) Cisplatin and PIC (5 mg/kg) treated group, (F) Cisplatin and PIC (10 mg/kg) treated group and (G) Cisplatin and PIC (20 mg/kg) treated group

(A) and (B) showed a normal structure of glomeruli and tubules. (C) and (D) showed cisplatin-induced degeneration in tubular epithelial lining, hemorrhage, luminal dilatation and eosinophilic casts formation. (E) Cisplatin and PIC (5 mg/kg) co-treated group showed mild reduction in cisplatin-induced pathological alterations. Tubular necrosis and eosinophilic casts formation were observed. (F) and (G)

showed that concomitant treatment of rats with PIC at higher doses (10 and 20 mg/kg) ameliorated the pathological changes induced by cisplatin injection and restored the normal histological structure of the glomeruli and tubules. Mild focal hemorrhage in the cortex was detected in the kidney specimens from the group co-treated with PIC (10 mg/kg) (F).

Table 2: Aspects of histopathological alterations in kidney specimens

| Group | Degeneration | Necrosis | Hemorrhage | Casts formation |
|--------------------------|--------------|----------|------------|-----------------|
| Control | - | - | - | - |
| Cisplatin | +++ | ++ | +++ | +++ |
| Cisplatin+PIC (5 mg/kg) | + | + | - | ++ |
| Cisplatin+PIC (10 mg/kg) | - | - | + | - |
| Cisplatin+PIC (20 mg/kg) | - | - | - | - |
| PIC (20 mg/kg) | - | - | - | - |

+++ (severe), ++ (moderate), + (mild), - (nil), PIC: Piceatannol.

Effect of PIC on oxidative stress markers

Cisplatin significantly depleted GSH by 64% and increased MDA levels by 40%, as compared to control group. In contrast, pre- and co-treatment of rats with the most effective dose of PIC (10 mg/kg)

showed a significant increase in GSH level by 214% and a significant decrease in MDA level by 21%, as compared to cisplatin group. Animals treated with PIC alone did not show any significant alterations in both GSH and MDA levels, as compared to the control group (fig. 3A and 3B).

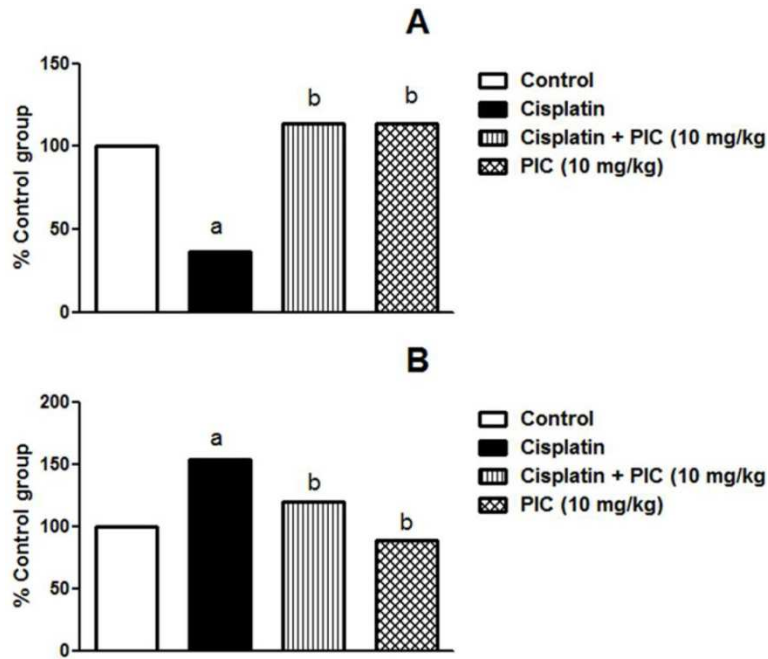


Fig. 3: Effect of piceatannol (PIC) on oxidative stress markers in cisplatin-treated rats. (A)Reduced glutathione level (GSH). (B) Lipid peroxidation expressed a malondialdehyde level (MDA), Data were presented as % of control. a or b: significantly different from the control or cisplatin group respectively at P<0.05 using ANOVA followed by Tukey-Kramer as a post-hoc test

Effect of PIC on NF-κB (p65) expression

The expression of the inflammatory marker NF-κB was estimated using immunohistochemical staining. The control group showed minimal immunostaining. No change in NF-κB expression was observed in the group treated with PIC alone. Cisplatin treatment

increased the expression of NF-κB, as shown by the intense brown staining. Concomitant treatment with PIC (10 mg/kg) significantly mitigated cisplatin-induced expression of NF-κB. The immunohistochemical staining of NF-κB was quantified as the optical density of the stained regions using the image analysis software (fig. 4).

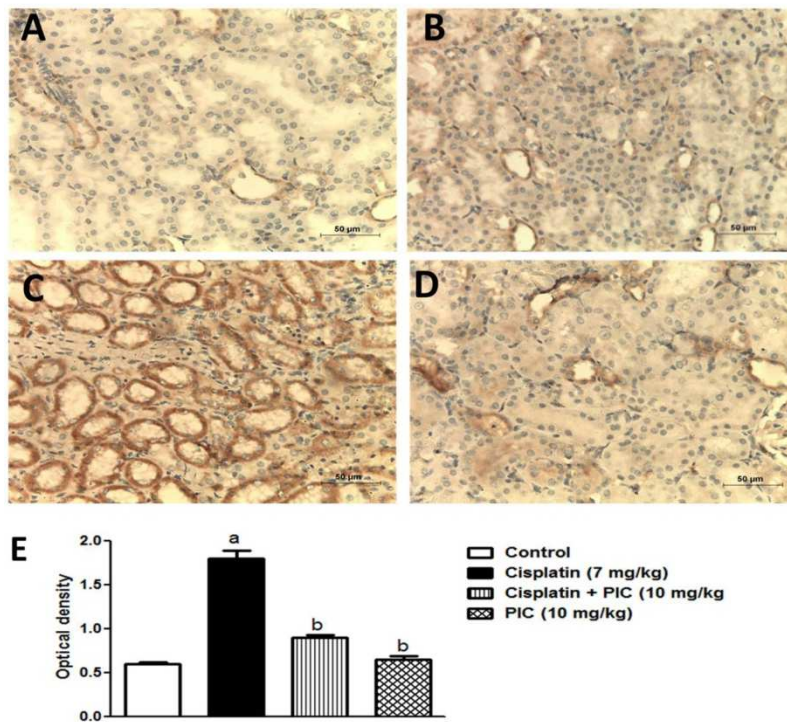


Fig. 4: Effect of piceatannol (PIC) NF-κB (p65) expression in cisplatin-treated rats, Control (A) and PIC alone (B) groups: negative immunostaining, Cisplatin (C): extensive expression, Cisplatin+PIC (D): mild immunostaining. (E): Quantitative image analysis for immunohistochemical staining were expressed as optical densities across 10 different fields for each section. Data were presented as means±SD (n=8). a or b: significantly different from the control or cisplatin group respectively, at P<0.05 using ANOVA followed by Tukey-Kramer as a post-hoc test

DISCUSSION

Cisplatin is a highly effective chemotherapeutic agent, used for the treatment of a wide spectrum of solid tumors but unfortunately, it causes dose-dependent nephrotoxicity that may lead to dose reduction or withdrawal. Nowadays, the usage of natural products as a protective strategy against cisplatin-induced nephrotoxicity represents a great scope of pharmacological research [22]. The current study provides an evidence of the protective effect of PIC against cisplatin-induced nephrotoxicity.

A dose-response study was conducted to choose the optimal dose of PIC for further investigations. Cisplatin was given i. p. as a single dose (7 mg/kg) causing significant alterations in nephrotoxicity markers; an increase in serum creatinine, BUN and relative kidney weight, as well as a decrease in total body weight. Increased mortality rate was also observed. The aforementioned abnormalities and biochemical changes were further confirmed by histological examination. It is well documented that cisplatin penetrates the tubular cells and reaches high concentrations in the proximal tubular cells of the inner renal cortex and outer medulla (S3 segment). Therefore, these areas are the major sites for cisplatin-induced renal injury, which in turn, leads to injury of other tubular areas including the distal and collecting tubules [23]. In the present study, cisplatin treatment caused severe nephrotoxic manifestations including degeneration of cortical epithelium, congestion, hemorrhage and necrosis, as well as eosinophilic casts formation in the lumen of proximal and distal convoluted tubules. Proximal convoluted tubules showed necrosis and degeneration of epithelial lining cells. These results were in agreement with previous studies [24-27].

Treatment of rats with 10 mg/kg PIC i. p. for 7 d, starting two days before cisplatin single dose, significantly ameliorated the aforementioned cisplatin-induced alterations in kidney, as compared to the lowest dose (5 mg/kg). The highest dose (20 mg/kg) showed no significant benefit over the middle dose (10 mg/kg). In addition, histological examination confirmed the previous results. Based on these data, PIC in the dose 10 mg/kg has proven efficacy in protecting against cisplatin-induced nephrotoxicity and was used to assess the possible mechanisms underlying this nephroprotective effect.

Oxidative stress is highly involved in the pathogenesis of cisplatin-induced kidney injury [28]. Cisplatin induces ROS production in renal epithelial cells, mainly by decreasing the activity of antioxidant enzymes and by GSH depletion which directly act on cellular components, including lipids, proteins and DNA which affect their structure and function [29]. In the present study, cisplatin injection (7 mg/kg) induced oxidative stress demonstrated by a significant depletion of reduced glutathione level and significant increase of lipid peroxidation in renal tissues, as compared to control group. These results were in agreement with previous studies [30-32]. In contrast, PIC restored reduced glutathione to normal levels and significantly decreased the extent of lipid peroxidation. The antioxidant effect of PIC was proven in previous studies [33, 34].

Inflammation plays a major role in the pathogenesis of cisplatin-induced nephrotoxicity, where a myriad of proinflammatory cytokines and chemokines are induced [1]. NF- κ B can mediate inflammatory response, by affecting the expression of a large number of proinflammatory mediators such as TNF- α , COX-2 and inducible nitric oxide [35]. Cisplatin-induced ROS production activates NF- κ B, which in turn induces the production of proinflammatory cytokines [36]. In the current study, the cisplatin-treated group showed a pro-inflammatory response as evidenced by significant increase in NF- κ B expression. In contrast, concomitant treatment with PIC significantly decreased NF- κ B expression and hence inhibited the downstream inflammatory cascade. In this regard, studies have shown that PIC could suppress NF- κ B activation by different inflammatory inducers such as H₂O₂, lipopolysaccharides (LPS), okadaic acid, and ceramide [14]. In addition, our results were in accordance with previous studies that have reported the anti-inflammatory effect of PIC and its ability to diminish the production of inflammatory cytokines [15, 37].

CONCLUSION

These findings indicate for the first time that PIC may have a potential protective effect against cisplatin-induced renal injury, by restoring

creatinine and BUN to normal levels and reversing the histopathological alterations in the kidney. A dose 10 mg/kg was the most effective and demonstrated an antioxidant effect against cisplatin-induced oxidative stress. Moreover, PIC significantly decreased NF- κ B expression. Further studies are required to determine the possible underlying mechanisms of the nephroprotective effect of PIC, in terms of oxidative, inflammatory and apoptotic markers.

CONFLICT OF INTERESTS

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

REFERENCES

1. Miller RP, Tadagavadi RK, Ramesh G, Reeves WB. Mechanisms of cisplatin nephrotoxicity. *Toxins (Basel)* 2010;2:2490-518.
2. Santos NA, Catão CS, Martins NM, Curti C, Bianchi ML, Santos AC. Cisplatin-induced nephrotoxicity is associated with oxidative stress, redox state unbalance, impairment of energetic metabolism and apoptosis in rat kidney mitochondria. *Arch Toxicol* 2007;81:495-504.
3. Arany I, Safirstein RL. Cisplatin nephrotoxicity. *Semin Nephrol* 2003;23:460-4.
4. Peres LA, da Cunha AD Jr. Acute nephrotoxicity of cisplatin: molecular mechanism. *J Bras Nefrol* 2013;35:332-40.
5. Sánchez-González PD, López-Hernández FJ, López-Novoa JM, Morales AI. An integrative view of the pathophysiological events leading to cisplatin nephrotoxicity. *Crit Rev Toxicol* 2011;41:803-21.
6. Ozkok A, Edelstein CL. Pathophysiology of cisplatin-induced acute kidney injury. *BioMed Res Int* 2014;1:1-17.
7. Khan SA, Priyamvada S, Khan W, Khan S, Farooq N, Yusufi AN. Studies on the protective effect of green tea against cisplatin induced nephrotoxicity. *Pharmacol Res* 2009;60:382-91.
8. Sindhu G, Nishanthi E, Sharmila R. Nephroprotective effect of vanillic acid against cisplatin induced nephrotoxicity in wistar rats: a biochemical and molecular study. *Env Toxicol Pharmacol* 2015;39:392-404.
9. Piotrowska H, Kucinska M, Murias M. Biological activity of piceatannol: leaving the shadow of resveratrol. *Mutat Res* 2012;750:60-82.
10. Ferrigni NR, McLaughlin JL, Powell RG, Smith Jr CR. Use of potato disc and brine shrimp bioassays to detect activity and isolate piceatannol as the antileukemic principle from the seeds of *Euphorbia lagascae*. *J Natural Prod* 1984;47:347-52.
11. Tyagi S, Chirag JP, Raghvendra. Biological and clinical spectrum of piceatannol-a hydroxylated analogue of resveratrol: a phytochemical review. *J Biomed Pharm Res* 2012;1:1-5.
12. Kukreja A, Wadhwa N, Tiwari A. Therapeutic role of resveratrol and piceatannol in disease prevention. *J Blood Disorders Transf* 2014;5:1-6.
13. Rossi M, Caruso F, Opazo C, Saliccioli J. Crystal and molecular structure of piceatannol; scavenging features of resveratrol and piceatannol on hydroxyl and peroxy radicals and docking with transthyretin. *J Agric Food Chem* 2008;56:10557-66.
14. Ashikawa K, Majumdar S, Banerjee S, Bharti AC, Shishodia S, Aggarwal BB. Piceatannol inhibits TNF-Induced NF- κ B activation and NF- κ B-mediated gene expression through suppression of I κ B α Kinase and p65 phosphorylation. *J Immunol* 2002;169:6490-7.
15. Youn J, Lee JS, Na HK, Kundu JK, Surh YJ. Resveratrol and piceatannol inhibit iNOS expression and NF- κ B activation in dextran sulfate sodium-induced mouse colitis. *Nutr Cancer* 2009;61:847-54.
16. Kalariya NM, Shoebc M, Aramati BM, Reddy ABM, Sawhney R, Ramana KV. Piceatannol suppresses endotoxin-induced ocular inflammation in rats. *Int Immunopharmacol* 2013;17:439-46.
17. Son PS, Park SA, Na HK, Jue DM, Kim S, Surh YJ. Piceatannol, a catechol-type polyphenol, inhibits phorbol ester-induced NF- κ B activation and cyclooxygenase-2 expression in human breast epithelial cells: cysteine 179 of IKK β as a potential target. *Carcinogenesis* 2010;31:1442-9.
18. Farrand L, Byun S, Kim JY, Im-Aram A, Lee J, Lim S, et al. Piceatannol enhances cisplatin sensitivity in ovarian cancer via modulation of

- p53, X-linked Inhibitor of apoptosis protein (XIAP), and mitochondrial fission. *J Biol Chem* 2013;288:23740-50.
19. Butler AR. The Jaffé reaction. Part II. A kinetic study of the Janovsky complexes formed from creatinine (2-imino-1-methylimidazolidin-4-one) and acetone. *J Chem Soc Perkin Trans* 1975;2:853-7.
 20. Clark VL, Kruse JA. Clinical methods: the history, physical, and laboratory examinations. *JAMA* 1990;264:2808-9.
 21. Bancroft JD, Stevens A, Turner DR. Theory and practice of histological techniques. 4th ed. Churchill Livingstone, New York; 1996.
 22. Park JY, Choi P, Kim T, Ko H, Kim HK, Kang KS, *et al.* Protective effects of processed ginseng and its active ginsenosides on cisplatin-induced nephrotoxicity: *in vitro* and *in vivo* studies. *J Agric Food Chem* 2015;63:5964-9.
 23. Taguchi T, Nazneen A, Abid MR, Razaque MS. Cisplatin associated nephrotoxicity and pathological events. *Contrib Nephrol Basel, Karger* 2005;148:107-21.
 24. Gautam RK, Singh RK, Ms K. Evaluation of nephroprotective activity of mentha arvensis in cisplatin induced nephrotoxicity. *Asian J Pharm Clin Res* 2014;7:188-91.
 25. Saleh S, Ain-Shoka AA, El-Demerdash E, Khalef MM. Protective effects of the angiotensin II receptor blocker losartan on cisplatin-induced kidney injury. *Chemotherapy* 2009;55:399-406.
 26. Arjumand W, Sultana S. Glycyrrhizic acid: a phytochemical with a protective role against cisplatin-induced genotoxicity and nephrotoxicity. *Life Sci* 2011;89:422-9.
 27. Reddy UK, Kumar S. Protective effect of *Abutilon indicum* L. (Malvaceae) against cisplatin induced nephrotoxicity in rats. *Innovare J Life Sci* 2013;17:35-9.
 28. Yao X, Panichpisal K, Kurtzman N, Nugent K. Cisplatin nephrotoxicity: a review. *Am J Med Sci* 2007;334:115-24.
 29. Huang Q, Dunn RT, Jayadev S, DiSorbo O, Pack FD, Farr SB, *et al.* Assessment of cisplatin induced nephrotoxicity by microarray technology. *Toxicol Sci* 2001;63:196-207.
 30. Sahu BD, Kumar JM, Sistla R. Baicalein, a bioflavonoid, prevents cisplatin-induced acute kidney injury by up-regulating antioxidant defenses and down-regulating the MAPKs and NF- κ B pathways. *PLoS One* 2015;10:1-19.
 31. Praharshta AM, Samuel V, Nirmala P. Role of ebselen, a selenoorganocompound in cisplatin induced nephrotoxicity in wistar rats. *Int J Chem Pharm Res* 2015;7:57-9.
 32. Mondy S, Kvsrg P, Jhansi D, Vijay R. Prophylactic and curative effect of ethanolic extract of *Bassia malabarica* bark against cisplatin induced nephrotoxicity. *Asian J Pharm Clin Res* 2014;7:143-6.
 33. Rhayem Y, Thérond P, Camont L, Couturier M, Beaudoux JL, Legrand A, *et al.* Chain-breaking activity of resveratrol and piceatannol in a linoleate micellar model. *Chem Phys Lipids* 2008;155:48-56.
 34. Woo A, Min B, Ryou S. Piceatannol-3'-O- β -D-glucopyranoside as an active component of rhubarb activates endothelial nitric oxide synthase through inhibition of arginase activity. *Exp Mol Med* 2010;42:524-32.
 35. Abraham E. Nuclear factor- κ B and its role in sepsis-associated organ failure. *J Infect Dis* 2003;187 Suppl 2:364-9.
 36. Oh GS, Kim HJ, Shen AH, Lee SB, Yang SH, Shim H, *et al.* New therapeutic concept of NAD redox balance for cisplatin nephrotoxicity. *Biomed Res Int* 2016;1:1-12.
 37. Jin CY, Moon DO, Lee KJ, Kim MO, Lee JD, Choi YH. Piceatannol attenuates lipopolysaccharide-induced NF-kappaB activation and NF-kappaB-related proinflammatory mediators in BV2 microglia. *Pharmacol Res* 2006;54:461-7.

How to cite this article

- Sara A Wahdan, Samar S Azab, Doaa A Elsherbiny, Ebtehal EL Demerdash. Piceatannol ameliorates cisplatin-induced histological and biochemical alterations in rats kidney. *Int J Pharm Pharm Sci* 2017;9(10):305-311.