

IN VITRO ANTIOXIDANT, ANTIMICROBIAL AND ADMET STUDY OF NOVEL FURAN/BENZOFURAN C-2 COUPLED QUINOLINE HYBRIDS

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ABSTRACT

Objective: Synthesis of novel 2-(benzofuran-2-yl) and 2-(furan-2-yl) quinoline-4-carboxylates and their [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol, [2-(1-furan-2-yl) quinolin-4-yl] methanol and its derivatives for antioxidant, antimicrobial and *in silico* pharmacokinetic study.

Methods: Synthesis was carried with the conventional method and the structures were confirmed by IR, ¹H NMR, ¹³C NMR and mass spectral analysis. The antioxidant activity was performed by DPPH and H₂O₂ radical scavenging method. The antimicrobial investigation was established by cup plate and food poison technique. The *in silico* absorption, distribution, metabolism, excretion and toxicity (ADMET) study of the drug was carried out in ACD/lab-2.

Results: The antioxidant activity results revealed that compounds 4b-c, 5a-b, 10c and 10f exhibited good DPPH radical and hydrogen peroxide scavenging activity. The antibacterial results revealed that compounds 4c, 5a-b, 10b, 10d and 10f exhibited good activity against *Escherichia coli*, *Klebsiella pneumonia* and *Salmonella typhimurium*. Further, the antifungal activity results showed that compounds 4c, 5c and 10c-e were showing good activity against *Aspergillus flavus* and *Candida neoformans*. The mean value of P<0.05 were considered to be statistically significant. The ADMET results revealed that compounds emerged as a potential candidate for antioxidant and antimicrobial agents.

Conclusion: The study reveals that compounds containing furan/benzofuran coupled heterocycles played the important role for activity as they possess potent antioxidant and antimicrobial agents. The *in silico* ADME analysis also suggesting the compounds were in acceptable range to obey the pharmacokinetic parameters.

Keywords: DPPH, Lipinski, ADMET, Toxicity, *Escherichia coli* and *Aspergillus flavus*

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INTRODUCTION

Free radicals are species capable of free existence that contains one or more unpaired electrons which react with another molecule by accepting or donating the electrons [1]. The harmful intervention of reactive oxygen species (ROS) in normal metabolic processes leads to pathologic changes which is a consequence of their interaction with biomolecules inside and outside the cells [2, 3]. ROS contain molecules like hydrogen peroxide (H₂O₂); hydroxyl radical (-OH) and superoxide (Oz-), by the generation of ROS, there is an alteration in the normal functioning of the cell and leads to pathophysiological changes. Free radicals are responsible for causing a wide number of health hazards such as cancer, ageing, heart diseases and gastric problems, etc. Oxygen free radicals disintegrate DNA and destroy cell membranes by enzymatic metabolic processes [4-6]. In nature's collection of biologically active molecules, benzofuran derivatives constitute a major group [7-9]. Benzofuran ring systems bearing substitutions at the C-2 position are widely distributed in nature and have been reported to have antioxidant, antiviral, antifungal activities [10, 11], antimicrobial [12, 13], anti-inflammatory [14], antipsychotic [15], analgesic [16], antilipidemic [17] and CNS stimulant activities [18]. Investigation of benzofuran derivatives in search of new drugs or to increase the efficacy of the present drugs has been most frequent approach [19]. Similarly, furan and its analogues were found to be biologically useful. 2-(furan-2-yl) quinoline-4-carboxylic acid and its analogues were reported to have the inhibiting property of *C. albicans* prolyl-tRNA synthetase and showed potent *in vitro* antifungal activities against dermatophytes [20, 21], 2-(furan-2-yl)-4-(phenoxy) quinoline derivatives were screened for cytotoxicity and anti-inflammatory activities [22], 4-

(phenylamino) furo-[2, 3-b] quinoline and 2-(furan-2-yl)-4-(phenylamino) quinoline derivatives for cytotoxicity evaluation and 1-{4-[(furo[2, 3-b] quinoline-4-yl) amino] phenyl} ethanone exhibited potent and broad spectrum of cytotoxicity [23, 24]. The quinoline ring system is an essential structural fragment of a large number of natural and synthesized compounds displaying interesting biological activities such as antimalarial, antibacterial, anti-asthmatic, antihypertensive, and anti-inflammatory [25-27]. Quinolines and their derivatives have been found applications as pharmaceuticals and agrochemicals, as well as being general synthetic building blocks [28-31]. Molecular hybridization is a rational approach to design new prototypes after coupling different pharmacophoric subunits that can be recognized by two or more biologic receptors [32]. These strategies have been used in drug discovery to increase the efficacy.

In view of these observations and in continuation of our work on drug discovery through cinchophene and their derivatives [33-36], herein we report a facile, inexpensive procedure in the preparation of novel hybrid molecules [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol and [2-(1-furan-2-yl) quinolin-4-yl] methanol derivatives using mild conditions, and explored for their *in vitro* antioxidant, antimicrobial and preliminary *in silico* ADMET properties.

MATERIALS AND METHODS

Materials

Chemicals used in the synthesis of compounds were from Alfa Aesar, Pvt. Ltd. Bangalore, India and Spectrochem Pvt. Ltd. Bangalore, India. The solvents were of reagent grade and when necessary, they

were purified and dried. Melting points of the synthesized compounds were determined with the help of Raga digital melting point apparatus and are uncorrected; Infrared data were recorded on a Bruker spectrophotometer using KBr pellets. ^1H and ^{13}C NMR spectra were recorded on Bruker AVANCE II 400 and 100 MHz instruments using $\text{DMSO-}d_6/\text{CDCl}_3$ as a solvent and TMS as an internal standard; chemical shifts are expressed as δ values (ppm). The J values are expressed in Hertz (Hz). Mass spectra (MS) were recorded in JEOL GCMATE II LC-Mass spectrometer with electron impact ionization (EI) technique. Analytical thin-layer chromatography (TLC) was performed with precoated TLC sheets of silica gel 60 F254 (Merck, Darmstadt, Germany), visualized by long and short wavelength UV lamps (nm). Chromatographic purifications were performed on Merck silica gel (100-200 mesh).

Methods

Synthesis of 1-(1-benzofuran-2-yl) ethanone(1)

Synthesis of 1-(1-benzofuran-2-yl) ethanone was achieved by the addition of salicylaldehyde (5.8 g, 0.047 mol), chloroacetone (4.3g, 0.047 mol) to an alcoholic KOH (33 %, 20 ml) solution and kept for stirring vigorously for about 2-3 h, maintaining a temperature 0-5 °C. The resultant mixture was poured onto crushed ice. The separated solid was filtered and recrystallized from petroleum ether (60-80). The yield was 85 %, M. Pt. 75-78 °C.

General procedure for the synthesis of substituted 2-(1-benzofuran-2-yl) quinoline-4-carboxylic acid 3(a-c)

A mixture of 1-(1-benzofuran-2-yl) ethanone (1.8 g, 0.0113 mol) and substituted 1H-indole-2, 3-dione (1.5 g, 0.0113 mol) in ethanol (10 ml) and aqueous solution of KOH (33 %) 5 ml was added, the reaction mixture was stirred at 65-70 °C for about 12-14 h. The reaction mixture was extracted with ethyl acetate (2-3 times) and the aqueous layer was poured onto crushed ice, acidified with 10 M HCl and the resulting mass was filtered and dried to get yellow amorphous powder, yield 80 %.

General procedure for the synthesis of substituted methyl 2-(1-benzofuran-2-yl) quinoline-4-carboxylates 4(a-c)

Analogues of 2-(1-benzofuran-2-yl) quinoline-4-carboxylic acids were dissolved in sufficient quantity of methanol with a catalytic amount of $\text{Conc. H}_2\text{SO}_4$ and the mixture were refluxed for about 10-12 h. The reaction mixture was cooled to room temperature and poured onto the crushed ice, filtered, washed with water, dried and recrystallized from petroleum ether (60-80) and ethyl acetate (3:1v/v).

General procedure for the synthesis of substituted [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol 5(a-c)

To a 100 ml round bottom flask, dissolve methyl 2-(1-benzofuran-2-yl) quinoline-4-carboxylate in sufficient quantity of ethanol and maintain the reaction mixture below 5 °C, followed by the addition of sodium borotetrahydride until effervescence ceases. Stir the reaction mixture overnight at room temperature and pour onto the crushed ice after neutralizing with 10 M HCl. The solid mass obtained was filtered, washed with water, dried and recrystallized from ethyl alcohol.

General procedure for the synthesis of substituted 2-(1-furan-2-yl) quinoline-4-carboxylic acid 8(a-f)

The compounds 8 (a-f) was synthesized by literature method [37] with slight modification. After completion of the reaction, the reaction mixture was chilled in a nice bath. The solid mass of the sodium salt of cinchonic acid was collected by filtration. The residue was dissolved in water and acidified with 10 % glacial acetic acid, filter the residue and repeatedly washed with ethyl acetate (4-5 times).

General procedure for the synthesis of substituted methyl 2-(1-furan-2-yl) quinoline-4-carboxylates 9(a-c)

Analogues of 2-(1-furan-2-yl) quinoline-4-carboxylic acids were dissolved in sufficient quantities of methanol with a catalytic amount of $\text{Conc. H}_2\text{SO}_4$ and was refluxed for about 12-14 h. The progress of the reaction was tartan by TLC. After completion, the reaction

mixture was cooled to room temperature and poured onto the crushed ice. The resulting solid mass was filtered, washed with water, dried and recrystallized from petroleum ether (60-80).

General procedure for the synthesis of substituted [2-(1-furan-2-yl) quinolin-4-yl] methanol 10(a-f)

The synthesis of substituted [2-(1-furan-2-yl) quinolin-4-yl] methanol 10(a-f) is similarly to the synthesis of analogues 5(a-c).

Spectral details

[2-(1-benzofuran-2-yl) quinolin-4-yl] methanol (5a)

Yield: 78 %. M. Pt. 154-156 °C; IR (KBr) ν_{max} 3114 (-O-H stretching), 3062, 1665, 1595, 1253 cm^{-1} ; ^1H NMR ($\text{DMSO-}d_6$, 400 MHz, δ ppm): 8.20 (1H, d, $J=8$ Hz, H-13), 8.10 (1H, s, H-19), 7.87 (1H, d, $J=12$ Hz, H-16), 7.58 (4H, m, H-9, 8, 7, 15), 7.50 (1H, t, $J=8$ Hz, H-6), 7.353 (1H, t, $J=1.2$ Hz, H-14), 7.260 (1H, t, $J=2.4$ Hz, H-3), 5.25 (2H, s, H-20), 1.72 (1H, s, quinolin-4-yl) methanol-OH proton); ^{13}C NMR ($\text{DMSO-}d_6$, 100 MHz, δ ppm): 159.1 (C, C-18), 157.1 (C, C-2), 156.6 (C, C-10), 154.5 (C, C-5), 154.0 (CH, C-13), 145.8 (CH, C-14), 141.2 (C, C-12), 139.6 (C, C-15), 123.6 (CH, C-16), 123.2 (CH, C-8), 122.6 (CH, C-9), 122.1 (CH, C-7), 121.5 (C, C-4), 115.8 (C, C-17), 114.9 (CH, C-19), 113.9 (CH, C-6), 108.5 (CH, C-3), 67.0 (CH_2 , C-20). Calcd. 275.3 gm/ml. EI-MS (m/z): 276.0 (M+1).

[2-(1-benzofuran-2-yl)-6-chloroquinolin-4-yl] methanol (5b)

Yield: 81 %. M. Pt. 206-208 °C; IR (KBr) ν_{max} 3120 (-O-H stretching), 2920, 1605, 1260 cm^{-1} ; ^1H NMR ($\text{DMSO-}d_6$, 400 MHz, δ ppm): 8.16 (1H, d, $J=4$ Hz, H-13), 8.14 (1H, s, H-19), 7.91 (1H, d, $J=4$ Hz, H-8), 7.68 (1H, t, $J=4$ Hz, H-8), 7.66 (1H, d, $J=4$ Hz, H-14), 7.653 (1H, d, $J=0.4$ Hz, H-7), 7.63 (1H, s, H-3), 7.37 (1H, t, $J=4$ Hz, H-9), 7.314 (1H, d, $J=0.8$ Hz, H-6), 5.23 (2H, s, H-21), 1.55 (1H, s, quinolin-4-yl) methanol-OH proton); ^{13}C NMR ($\text{DMSO-}d_6$, 100 MHz, δ ppm): 154.9 (C, C-18), 154.5 (C, C-5), 148.7 (C, C-2), 148.4 (C, C-10), 145.8 (C, C-12), 131.4 (C, C-15), 131.2 (CH, C-14), 130.4 (CH, C-13), 128.6 (CH, C-7), 125.9 (C, C-4), 125.7 (C, C-17), 123.5 (CH, C-16), 122.7 (CH, C-8), 122.1 (CH, C-19), 115.5 (CH, C-9), 111.5 (CH, C-6), 106.6 (CH, C-3), 59.7 (CH_2 , C-21). Calcd. 309.30 gm/ml. EI-MS (m/z): 276.0 (M+1).

[2-(1-benzofuran-2-yl)-8-fluoroquinolin-4-yl] methanol (5c)

Yield: 79 %. M. Pt. 158-160 °C; IR (KBr) ν_{max} 3114 (-O-H stretching), 3063, 1597, 1253 cm^{-1} ; ^1H NMR ($\text{DMSO-}d_6$, 400 MHz, δ ppm): 8.20 (1H, d, $J=8$ Hz, H-16), 8.12 (1H, s, H-19), 7.89 (1H, d, $J=8$ Hz, H-6), 7.62 (3H, m, H-15, 9, 7), 7.51 (1H, t, $J=8$ Hz, H-8), 7.35 (1H, t, $J=8$ Hz, H-14), 7.26 (1H, t, $J=8$ Hz, H-3), 5.26 (2H, s, H-21), 1.65 (1H, s, quinolin-4-yl) methanol-OH proton); ^{13}C NMR ($\text{DMSO-}d_6$, 100 MHz, δ ppm): 154.9 (C, C-18), 154.8 (C, C-5), 149.1 (C, C-2), 148.0 (C, C-10), 147.2 (C, C-12), 129.9 (CH, C-15), 129.4 (CH, C-14), 128.3 (C, C-13), 126.6 (CH, C-7), 125.7 (C, C-4), 124.8 (C, C-17), 123.4 (CH, C-16), 122.0 (CH, C-8), 114.4 (CH, C-9), 111.5 (CH, C-6), 106.1 (CH, C-3), 59.6 (CH_2 , C-21). Calcd. 293.00 gm/ml.

[6-chloro-2-(furan-2-yl) quinolin-4-yl] methanol (10a)

Yield: 88 %. M. Pt. 148-150 °C; IR (KBr) ν_{max} 3200 (-O-H stretching), 2918, 1672, 1604, 1249, 748 cm^{-1} ; ^1H NMR ($\text{DMSO-}d_6$, 400 MHz, δ ppm): 8.05 (1H, d, $J=1.2$ Hz, H-9), 7.92 (1H, s, H-12), 7.86 (1H, s, H-15), 7.631 (2H, t, $J=0.8$ Hz, H-4, 3), 7.221 (1H, d, $J=7.2$ Hz, H-5), 6.590 (1H, m, H-10), 5.16 (2H, s, H-17), 1.68 (1H, s, quinolin-4-yl) methanol-OH proton); ^{13}C NMR ($\text{DMSO-}d_6$, 100 MHz, δ ppm): 152.9 (C, C-14), 148.6 (C, C-2), 148.4 (C, C-6), 145.8 (C, C-8), 131.1 (CH, C-5), 130.5 (C, C-11), 130.2 (CH, C-10), 125.3 (CH, C-9), 122.7 (C, C-13), 114.8 (CH, C-12), 112.6 (CH, C-15), 110.7 (CH, C-4), 59.7 (CH_2 , C-17). Calcd. 259.68 gm/mol. EI-MS (m/z): 260.0 (M+1).

[6-bromo-2-(furan-2-yl) quinolin-4-yl] methanol (10b)

Yield: 83 %. M. Pt. 151-153 °C; IR (KBr) ν_{max} 3215 (-O-H stretching), 2923, 1670, 1603, 1248 cm^{-1} ; ^1H NMR ($\text{DMSO-}d_6$, 400 MHz, δ ppm): 8.022 (1H, d, $J=0.7$ Hz, H-9), 7.98 (1H, d, $J=8$ Hz, H-15), 7.90 (1H, s, H-12), 7.740 (1H, d, $J=2.4$ Hz, H-5), 7.630 (1H, d, $J=1.2$ Hz, H-10), 7.21 (1H, d, $J=4$ Hz, H-3), 6.58 (1H, m, H-4), 5.14 (2H, s, H-17), 1.67 (1H, s, quinolin-4-yl) methanol-OH proton); ^{13}C NMR ($\text{DMSO-}d_6$, 100 MHz, δ ppm): 152.9 (C, C-14), 148.6 (C, C-2), 148.2 (C, C-6), 145.9 (CH, C-5), 132.7 (CH, C-10), 131.2 (CH, C-9), 125.9 (CH, C-12), 125.8 (CH, C-11),

119.1 (CH, C-15), 114.7 (C, C-13), 112.6 (CH, C-3), 110.8 (CH, C-4), 59.7 (CH₂, C-17). Calcd. 304.13 gm/ml. EI-MS (*m/z*): 304.13 (M⁺).

[8-fluoro-2-(furan-2-yl) quinolin-4-yl] methanol (10c)

Yield: 82 %. M. Pt. 85-87 °C; IR (KBr) ν_{\max} 3112 (-O-H stretching), 2921, 1603, 1250 cm⁻¹; ¹H NMR (DMSO-*d*₆, 400 MHz, δ ppm): 7.89 (1H, s, H-15), 7.60 (2H, t, *J*=4 Hz, H-11, 4), 7.36 (2H, d, *J*=4 Hz, H-10, 12), 6.58 (1H, d, *J*=4 Hz, H-3), 5.15 (2H, s, H-17), 1.66 (1H, s, quinolin-4-yl methanol-OH proton); ¹³C NMR (DMSO-*d*₆, 100 MHz, δ ppm): 158.6 (C, C-14), 156.1 (C, C-9), 153.0 (C, C-2), 149.0 (CH, C-5), 148.3 (C, C-8), 145.1 (CH, C-11), 137.3 (C, C-13), 126.1 (CH, C-12), 119.4 (CH, C-15), 114.5 (CH, C-10), 113.8 (C, C-6), 112.6 (CH, C-3), 110.9 (CH, C-4), 59.7 (CH₂, C-17). Calcd. 243.23 gm/ml. EI-MS (*m/z*): 244.10 (M+1).

[6-chloro-2-(5-methylfuran-2-yl) quinolin-4-yl] methanol (10d)

Yield: 77 %. M. Pt. 136-138 °C; IR (KBr) ν_{\max} 3186 (-O-H stretching), 2897, 1672, 1607, 1245, 752 cm⁻¹; ¹H NMR (DMSO-*d*₆, 400 MHz, δ ppm): 8.03 (1H, d, *J*=8 Hz, H-10), 7.83 (2H, d, *J*=4 Hz, H-11, 13), 7.59 (1H, d, *J*=4 Hz, H-16), 7.12 (1H, d, *J*=4 Hz, H-3), 6.191 (1H, d, *J*=0.8 Hz, H-4), 5.14 (2H, s, H-18), 2.46 (3H, s, H-6), 1.81 (1H, s, quinolin-4-yl methanol-OH proton); ¹³C NMR (DMSO-*d*₆, 100 MHz, δ ppm): 154.4 (C, C-15), 151.5 (C, C-7), 148.1 (C, C-5), 145.8 (C, C-2), 131.0 (C, C-9), 130.2 (C, C-12), 130.0 (CH, C-10), 125.1 (CH, C-11), 122.7 (C, C-14), 114.6 (CH, C-13), 112.1 (CH, C-16), 109.0 (CH, C-3, 4), 59.7 (CH₂, C-18), 13.6 (CH₃, C-6). Calcd. 273.71 gm/ml. EI-MS (*m/z*): 274.08 (M+1).

[6-bromo-2-(5-methylfuran-2-yl) quinolin-4-yl] methanol (10e)

Yield: 81 %. M. Pt. 104-106 °C; IR (KBr) ν_{\max} 3206 (-O-H stretching), 2922, 1667, 1535, 1245 cm⁻¹; ¹H NMR (DMSO-*d*₆, 400 MHz, δ ppm): 7.970 (1H, d, *J*=2 Hz, H-10), 7.93 (1H, d, *J*=8 Hz, H-11), 7.77 (1H, s, H-13), 7.701 (1H, d, *J*=2 Hz, H-16), 7.09 (1H, d, *J*=4 Hz, H-4), 6.181 (1H, d, *J*=0.8 Hz, H-4), 5.09 (2H, s, H-18), 2.45 (3H, s, H-6), 1.93 (1H, s, quinolin-4-yl methanol-OH proton); ¹³C NMR (DMSO-*d*₆, 100 MHz, δ ppm): 154.4 (C, C-15), 151.4 (C, C-5), 148.6 (C, C-7), 147.9 (C, C-2), 146.0 (C, C-9), 132.6 (C, C-12), 131.1 (CH, C-11), 125.8 (CH, C-10), 125.6 (C, C-14), 118.8 (CH, C-13), 114.6 (CH, C-16), 112.1 (CH, C-4), 109.0 (CH, C-3), 59.7 (CH₂, C-18), 13.5 (CH₃, C-6). Calcd. 318.16 gm/ml. EI-MS (*m/z*): 318.04 (M⁺).

[8-fluoro-2-(5-methylfuran-2-yl) quinolin-4-yl] methanol (10f)

Yield: 78 %. M. Pt. 118-120 °C; IR (KBr) ν_{\max} 3332 (-O-H stretching), 2922, 1599, 1247 cm⁻¹; ¹H NMR (DMSO-*d*₆, 400 MHz, δ ppm): 7.76 (1H, s, H-16), 7.52 (1H, t, *J*=4 Hz, H-12), 7.30 (2H, m, H-13, 3), 7.13 (1H, d, *J*=4 Hz, H-4), 6.16 (1H, d, *J*=4 Hz, H-11), 5.10 (2H, s, H-18), 2.42 (3H, s, H-6), 1.81 (1H, s, quinolin-4-yl methanol-OH proton); ¹³C NMR (DMSO-*d*₆, 100 MHz, δ ppm): 158.6 (C, C-15), 154.4 (C, C-7), 151.5 (C, C-5), 148.7 (C, C-9), 148.3 (C, C-2), 125.9 (CH, C-12), 125.5 (C, C-10), 119.3 (CH, C-11), 114.9 (C, C-14), 113.9 (CH, C-13), 113.7 (CH, C-16), 112.2 (CH, C-4), 109.0 (CH, C-3), 59.7 (CH₂, C-18), 13.6 (CH₃, C-6). Calcd. 257.25 gm/ml. EI-MS (*m/z*): 358.00 (M+1).

Pharmacokinetic parameter studies

ADME-toxicity prediction

The molecular descriptors of synthesized compounds (4a-c, 5a-c and 10a-f) are predicted by pharmacokinetics parameters like absorption, distribution, metabolism, excretion and toxicity (ADMET). The ADMET/SAR [38] helps to evaluate biologically active molecules and eliminate a biologically poor molecule, an active lead molecule which contains undesirable functional groups based on Lipinski rule. The statistical calculation for lead molecules includes surface area, geometry and fingerprint properties which help to understand biological important endpoints. Aqueous solubility (PlogS), blood-brain barrier penetration (QlogBB), intestinal absorption (logHIA) [39] and hepatotoxicity, Caco-2 cell permeability (QPPCaco) also help to predict the toxicity of lead molecules with intraperitoneal, oral, intravenous and subcutaneous toxic effects. The *in silico* study enables to decide the safety and efficacy of active molecules take up the molecule for in-depth studies.

Calculation of pharmacokinetic parameters and toxicity potential

Chemical structures and SMILES notations of the title compounds were obtained by using ACD labs Chem sketch version 12.0. SMILES

notations of the derivatives were then fed in the online free version 2011.06 to calculate various molecular properties and to predict the bioactivity score for drug targets including enzymes and nuclear receptors, kinase inhibitors, GPCR ligands and ion channel modulators. Molecular properties such as partition coefficient (Log P), topological polar surface area (TPSA), hydrogen bond donors and acceptors, rotatable bonds, number of atoms, molecular weight, and violations of Lipinski's rule of five were calculated to evaluate the drug-likeness of the synthesized compounds [40]. The bioactivity score and drug-likeness properties of the title compounds were compared with the standard drugs streptomycin, ciprofloxacin and pyrazinamide respectively.

Biological evaluation

DPPH free radical scavenging assay

The assay was performed after modification of the method described by Blois [41]. 0.1 ml of different concentrations of amethanolic solution of standard and test compounds (0.5, 0.75, 1.0, 1.25 and 1.5 mg/ml) was added to 2 ml of DPPH methanolic solution (60 mm). The mixture was shaken vigorously and allowed to react at room temperature and in darkness for 5 h. The absorbance of the resulting solution was measured at 517 nm using a UV/Vis spectrophotometer after 5 h incubation. Scavenging of DPPH free radicals was calculated as:

$$\text{DPPH scavenging activity (\%)} = \left[\frac{(\text{Ac}-\text{At})}{\text{Ac}} \right] \times 100$$

Where Ac is the absorbance of the control tube (containing all reagents except the test compound), and At is the absorbance of the test tube. Ascorbic acid was used as the standard in the concentration range of 0.5-1.5 mg/ml.

Hydrogen peroxide scavenging assay

H₂O₂ scavenging power was determined according to the method of Ruch and co-workers [42]. The method is based on the ability of a compound to convert hydrogen peroxide to water. 40 mM solution of hydrogen peroxide was prepared in saline phosphate buffer (pH 7.4). 100 μ l DMSO solutions of the test compounds or standards at the concentrations of 0.5, 0.75, 1.0, 1.25 and 1.5 mg/ml were separately added to 2 ml of the prepared hydrogen peroxide solution and the absorbance was measured at 230 nm after 10 min against a blank solution. The hydrogen peroxide scavenging activity for compounds and standards was calculated using the following equation

$$\text{H}_2\text{O}_2 \text{ scavenging activity (\%)} = \left[\frac{(\text{Ac}-\text{At})}{\text{Ac}} \right] \times 100$$

Where Ac is the absorbance of the control and At is the absorbance of the tested compounds or standards. Ascorbic acid at the concentration range of 0.5-1.5 mg/ml was used as the standard.

Antibacterial activity

Cup plate method

The antibacterial activity of synthesized molecules was studied systematically against three different strains of bacteria such as *E. coli* (ATCC No. 25922), *K. pneumonia* (ATCC No. 700603) and *S. typhimurium* (ATCC No. 14028) (gram-negative) by the agar diffusion method [43-45]. The organisms were subcultured using nutrient agar medium. The tubes containing sterilized medium were inoculated with respective bacterial strain. After incubation at 37 \pm 1 °C for 24 h. They were stored in a refrigerator. The flasks with incubated bacterial inoculums were prepared by transferring a loopful of stock culture to nutrient broth (100 ml) and incubated at 37 \pm 1 °C for 18 h. Before the experimentation, solutions of the test compounds were prepared by dissolving 2 ml each in dimethyl sulphoxide (5 ml). Reference standards for gram-negative bacteria were made by dissolving accurately weighed the quantity of Streptomycin respectively in dimethyl sulphoxide solution, separately. The nutrient agar medium was sterilized by autoclave at 121 °C (15 lb/sq. inch).

The petri-plates, test tubes and flasks containing medium is plugged with cotton were sterilized in hot air-oven at 160 °C, for an hrs. Into each sterilized pertained plate (10 cm diameter), about 20 ml each of molten nutrient bacteria (6 ml of inoculums to 300 ml of nutrient

agar medium) was transferred, aseptically. The plates were left at room temperature to allow for solidification. On each plate, four cups of 6 mm diameter were made with a sterile cork borer. Then, 0.1 ml of the test solution was added to the cups, aseptically and labelled, accordingly. The plates were kept undisturbed for at least 2 h. at room temperature to allow diffusion of the solution properly, into the nutrient agar medium. All the experiments were carried out in triplicate. Simultaneously, controls were maintained employing 0.1 ml of dimethyl sulphoxide to observe the solvent effects. After incubation of the plates at 37 ± 1 °C for 24 h, the diameter of the zone of inhibition was read with the help of antibiotic zone scale.

Antifungal activity

Poisoned food technique was performed to investigate the antifungal effect of test compounds against *Aspergillus flavus* and *Cryptococcus neoformans*. Synthesized compounds were tested for their antifungal activity [46]. The fungi employed for screening were sub-cultured using potato dextrose agar medium. The potato-dextrose-agar medium was sterilized by autoclave at 121 °C (15 lb/sq. inch), for 15 min. The petri-plates, tubes and flasks plugged with cotton were sterilized in an autoclave at 121 °C, for an hour. Into each sterilized petri-plate (10 cm diameter), about 30 ml each of molten potato dextrose-agar medium inoculated with the respective fungus (5 mm disc of the fungus grown) was transferred, aseptically. The petri dishes were incubated at 28 ± 1 °C temperature. The diameter of the zone of inhibition was read with the help of an 'antibiotic zone reader'. The experiments were performed in

triplicate in order to minimize the errors. The inhibition percentage of the *A. flavus* and *C. neoformans* mycelial growth was calculated using the following formula.

$$IP = 100 \times CT$$

Where C is the average of 3 replicates of mycelial growth (cm) of control petri dishes and T is the average of 3 replicates of mycelial growth (cm) of treated petri dishes. Fluconazole was used as the positive control and DMSO was thus used as the negative control.

RESULTS AND DISCUSSION

Chemistry

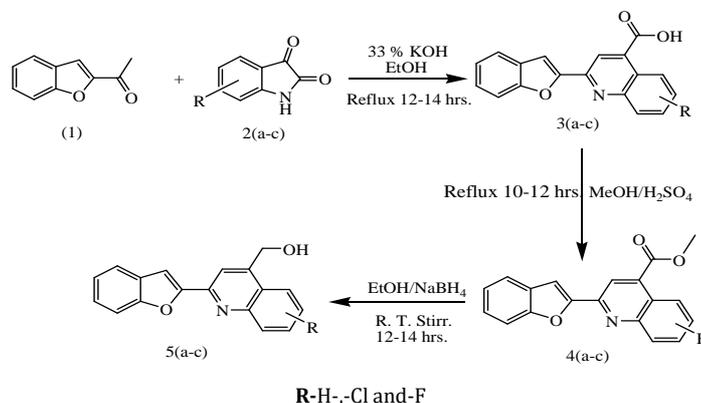
The synthesis of key intermediate 1-(1-benzofuran-2-yl) ethanone (1) and its utilization in the construction of 2-(1-benzofuran-2-yl) quinoline-4-carboxylic acids 3(a-c) and 8(a-f) are shown in Scheme I, II and table 1. The intermediate (1) was synthesized by stirring a mixture of 2-hydroxybenzaldehyde with chloroacetone in basic medium using methanol as solvent. Subsequently, 2-(1-benzofuran-2-yl) quinoline-4-carboxylic acids 3(a-c) were synthesized by the reaction of a compound (1) with substituted isatins 2(a-c) in basic medium to obtain the products 3(a-c). The ester, methyl 2-(1-benzofuran-2-yl) quinoline-4-carboxylates 4(a-c) were synthesized by reacting 3(a-c) with methyl alcohol in the presence of catalytic amount of Conc. H_2SO_4 to obtain the product in good yield. The alcohol 5(a-c) was synthesized by the reaction of compound 4(a-c) with reducing agent sodium borotetrahydride, under stirring condition.

Table 1: Particulars of the derivatives of [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol and [2-(1-furan-2-yl) quinolin-4-yl] methanol

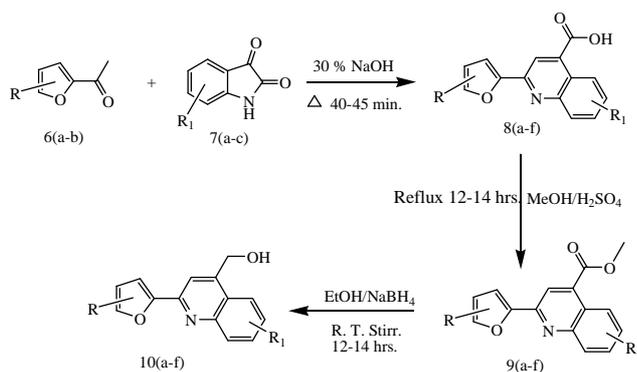
S. No.	Samples code	R	Molecular formula	Molecular weight	(%) yield	Melting point (°C)	
1	4a	-H-	$C_{19}H_{13}NO_3$	303.31	83	115-118	
2	4b	-Cl	$C_{19}H_{12}ClNO_3$	337.75	85	125-128	
3	4c	-F	$C_{19}H_{12}FNO_3$	301.30	80	122-124	
4	5a	-H-	$C_{18}H_{13}NO_2$	275.30	78	154-156	
5	5b	-Cl	$C_{18}H_{12}ClNO_2$	309.00	81	206-208	
6	5c	-F	$C_{18}H_{12}FNO_2$	293.00	79	158-160	
		R	R ₁				
7	10a	-H-	-Cl	$C_{14}H_{10}ClNO_2$	259.68	88	148-150
8	10b	-H-	-Br	$C_{14}H_{10}BrNO_2$	304.13	83	151-153
9	10c	-H-	-F	$C_{14}H_{10}FNO_2$	243.23	82	85-87
10	10d	-CH ₃	-Cl	$C_{15}H_{12}ClNO_2$	273.71	77	136-138
11	10e	-CH ₃	-Br	$C_{15}H_{12}BrNO_2$	318.16	81	104-106
12	10f	-CH ₃	-F	$C_{15}H_{12}FNO_2$	257.25	78	118-120

IR spectra of the compounds showed an absorption band in the range of 3114 cm^{-1} due to the characteristic-OH group stretching. The 1H NMR spectra of 5a showed a singlet at δ 5.25 ppm corresponding to methyl protons of [quinolin-4-yl] methanol, shown a singlet at δ 1.72 ppm corresponding to alcohol proton of methyl alcohol and the remaining peaks which appeared at δ 8.20 to 7.26 ppm corresponding to aromatic protons of the quinoline and benzofuran rings. The ^{13}C -NMR spectra of 5a showed a characteristic peak at δ 67.0 ppm corresponding to the methyl carbon of [quinolin-

4-yl] methanol, and the peaks between δ 159.1 to 108.5 ppm corresponding to aromatic carbons. MS analysis of 5(a-c) displayed their corresponding molecular ion peak confirming their molecular weight. IR spectra of the compounds showed the absorption band at 3215 cm^{-1} characteristic of-OH is stretching for different alcohols. The 1H NMR spectra of 10a showed a singlet at δ 5.16 ppm corresponding to methyl protons of [quinolin-4-yl] methanol, and another singlet at δ 1.68 ppm corresponding to the alcohol proton of [quinolin-4-yl] methanol.



Scheme I: Synthesis of substituted [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol 5(a-c)



R	R ₁
-H-	-Cl
-H-	-Br
-H-	-F
-CH ₃	-Cl
-CH ₃	-Br
-CH ₃	-F

Scheme II: Synthesis of substituted [2-(1-furan-2-yl)quinolin-4-yl] methanol 10(a-f)

The peaks between δ 8.08 to 6.59 ppm corresponding to benzofuran and quinoline ring protons. The ¹³C-NMR spectra of 10a showed a peak at δ 59.7 ppm corresponding to the methyl carbon of [quinolin-4-yl] methanol, and the peaks between δ 152.9 to 110.7 ppm corresponding to aromatic carbons. MS analysis of 10a displayed the molecular ion peak confirming their molecular weight. The ¹H NMR spectra of 10d showed a singlet at δ 5.14 ppm corresponding to methyl protons of [quinolin-4-yl] methanol, it showed singlet at δ 2.46 ppm corresponding to furan substituted methyl protons and appeared a singlet at δ 1.81 ppm corresponding to the alcohol proton of [quinolin-4-yl] methanol and the aromatic peaks which appeared between δ 8.05 to 6.19 ppm. The ¹³C-NMR spectra of 10d showed a peak at δ 59.7 ppm corresponding to the methyl carbon of [quinolin-4-yl] methanol, peak at δ 13.6 ppm corresponding to the methyl carbon of furan substituted methyl and the peaks between δ 154.4 to 109.0 ppm corresponding to aromatic carbons.

***In silico* ADMET (Absorption, distribution, metabolism, excretion and toxicity) profile**

The compounds with poor bioavailability show less effectiveness against disease. To overcome this problem, predicting bioavailability properties will be of great advantage in drug development. Hence, using computer-based methods like ADMET and SAR tools the molecular descriptors and drug likeliness properties was studied. The pharmacokinetic properties are represented in table 2 and 3. The *in silico* data of all the molecules displayed are within acceptable range. The interpretation of test compounds with reference standards (streptomycin, fluconazole and ascorbic acid) show that the compounds 4a-c, 5a-c and 10a-f were in

good, acceptable range and hence, further used to make an oral formulation for absorption and to transport proteins and metabolizing enzymes to maintain homeostatic condition. The intestinal absorption (\log_{HIA}) and Caco-2 cell permeability (PCaco-2) within the range of -2 poor absorption and +2 more absorption reveal that the compounds are more permeable in the intestine and helps for good transport of the drug or its metabolites.

The reference range of -5 (poor) to +1 (good) and substrate inhibitor from 0 to 1 in which the reference and test compounds 4a-c, 5a-c and 10a-f shows significant activity with human intestinal absorption and metabolism. The aqueous solubility of compounds lies with a range of 0 (poor) to 2 (good) showed that all the molecules have good solubility. While the reference compound, as well as test compounds, came within the acceptable range (table 3).

The toxicity of the substituted [2-(1-benzofuran-2-yl)quinolin-4-yl] methanol and [2-(1-furan-2-yl)quinolin-4-yl] methanol was predicted. The probability of health effects was predicted using ACD/I-Lab 2.0 (guest). The toxicity of selected compounds was listed in table 2. The LD₅₀ of potential compounds detects the cumulative potential of acute toxicity that is administered through oral, subcutaneous, intra peritoneal, intravenous and subcutaneous on mouse models. The comparative analysis of reference compounds with test compounds on subcutaneous, intra-peritoneal, oral and intravenous is low when compared to the reference molecule. The toxicity results suggest that the compounds 4a-c, 5a-c and 10a-f have less toxic effect to tissue and with no side effect (table 2). Hence, can be considered for further development.

Table 2: LD₅₀ ADME-TOX parameters and probability of health effects of substituted [2-(1-benzofuran-2-yl)quinolin-4-yl] methanol and [2-(1-furan-2-yl)quinolin-4-yl] methanol using ACD/I-Lab 2.0

Ligands	Intraperitoneal ^a	Oral ^b	Intravenous ^c	Subcutaneous ^d
4a	490(0.45)	540(0.15)	55(0.49)	490(0.28)
4b	500(0.4)	450(0.18)	56(0.48)	550(0.33)
4c	640(0.46)	460 (0.15)	58(0.47)	940(0.34)
5a	420(0.34)	840 (0.32)	46(0.47)	620(0.25)
5b	400(0.34)	810 (0.28)	42(0.47)	520(0.28)
5c	620(0.44)	690 (0.32)	47(0.47)	570(0.29)
10a	260(0.43)	800 (0.25)	65(0.4)	470(0.38)
10b	280(0.36)	870 (0.34)	67(0.35)	770(0.33)
10c	380(0.28)	550 (0.29)	78(0.38)	760(0.33)
10d	260(0.44)	780 (0.23)	62(0.39)	500(0.39)
10e	250(0.37)	840 (0.34)	59(0.35)	720(0.34)
10f	350(0.29)	580 (0.34)	67(0.38)	680(0.32)
Streptomycin	310(0.76)	880(0.53)	110(0.67)	400(0.52)
Fluconazole	1200(0.73)	1000(0.51)	580(0.47)	2700(0.23)
Ascorbic acid	1100(0.7)	4500(0.61)	820(0.58)	2700(0.5)

Estimated LD₅₀-mouse value in mg/kg after Intraperitoneal^a, Oral^b, Intravenous^c and Subcutaneous^d administration, The drugs with amoderate effect on reliability index (>0.5), The drugs with borderline effect on reliability index (>0.3,<0.5).

Table 3: ADME and pharmacological parameters prediction for the ligands substituted [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol and [2-(furan-2-yl) quinolin-4-yl] methanol using ADMET/SAR

Ligands	PlogBB ^a	PCaco ^b	logHIA ^c	logpGI (Non-substrate) ^d	logpGI (Non-inhibitor) ^e	PlogS ^f	logppapp ^g
4a	0.9430	0.5416	1.0000	0.7113	0.9010	-3.4558	1.0868
4b	0.9656	0.5853	1.0000	0.7231	0.8397	-4.3351	1.1113
4c	0.9709	0.5732	1.0000	0.7055	0.7329	-3.9795	1.1292
5a	0.9866	0.5175	1.0000	0.7710	0.7075	-2.8564	1.2208
5b	0.9906	0.5510	1.0000	0.7642	0.8692	-3.8706	1.2692
5c	0.9931	0.5465	1.0000	0.7690	0.8247	-3.4170	1.2863
10a	0.9906	0.5510	1.0000	0.7642	0.8692	-3.8706	1.2692
10b	0.9892	0.5277	1.0000	0.7637	0.8377	-3.6714	1.2573
10c	0.9931	0.5464	1.0000	0.7690	0.8247	-3.4170	1.2863
10d	0.9876	0.5621	1.0000	0.7598	0.8859	-3.7943	1.2395
10e	0.9855	0.5388	1.0000	0.7568	0.8570	-3.6238	1.2205
10f	0.9907	0.5559	1.0000	0.7609	0.8566	-3.3745	1.2440
Streptomycin	0.9712	0.8824	0.6968	0.5531	0.7577	-2.0122	-0.5128
Fluconazole	0.9382	0.9894	0.8867	0.6008	0.8782	-1.8626	1.3598
Ascorbic acid	0.8532	0.7710	0.6559	0.8696	0.9347	0.1081	-0.3148

^aPredicted blood/brain barrier partition coefficient (1-high penetration, 2-medium penetration and 3-low penetration). ^bpredicted Caco-2 cell permeability in nm/s (acceptable range-1 is poor,+1 is great). ^cpredicted human intestinal absorption in nm/s (acceptable range 0 is poor,>1 is great). ^dpredicted P-glycoprotein Substrate in nm/s (acceptable range of-5 is poor, 1 is great). ^epredicted P-glycoprotein inhibitor in nm/s (acceptable range 0-1). ^fpredicted aqueous solubility, (Concern value is 0-2 highly soluble). ^gpredicted Caco-2 cell Permeability in cm/s (Concern value is-1 to 1).

Drug-likeness score of the entitled compounds

Lipinski's rule of five is generally used by pharmaceutical chemists in drug design and development to predict bioavailability of lead or drug molecules. According to Lipinski's rule of five, a candidate molecule will likely be orally active, if: i) the calculated octanol/water partition coefficient (Log P)<5, ii) the molecular weight is under 500, iii) there were less than 5 hydrogen bond donors (OH and NH groups) and, iv) there are less than ten hydrogen bond acceptors (notably N and O). The molecular properties of [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol and [2-(1-furan-2-yl) quinolin-4-yl] methanol derivatives (4a-c, 5a-c and 10a-f) were calculated and are represented in table 4. Compounds (4a, 4c, 5a-c and 10a-f) did not violate any of the Lipinski's rules of five and were expected to be orally active. Molecular hydrophobicity indicated by Log P values for all the title compounds except 4b was found to be equal to 5 and it is in clear,

non-violation of Lipinski's rule of five, suggesting poor membrane permeability across the cell. Partition coefficient values of streptomycin, ciprofloxacin and pyrazinamide, the standard drugs were found to be well under 5 justifying their oral use. Molecular weight of twelve furan quinoline derivatives was found to be less than 500 and thus, molecules are likely to be easily transported, diffused and absorbed as compared to large molecules. A number of hydrogen bond acceptors (O and N atoms) and number of hydrogen bond donors (NH and OH) in the synthesized compounds 4a-c, 5a-c and 10a-f were in accordance with the Lipinski's rule of five i.e. less than 10 and 5, respectively. It can be predicted that the synthesized derivatives were likely to be orally active as they obeyed Lipinski's rule of five. The topological polar surface area is very much concurrently with the hydrogen bonding of a molecule and is a very good indicator bioavailability of drug molecules. TPSA of synthesized derivatives were observed in the range of 46.26-52.34 Å² and are well below the limit of 160 Å².

Table 4: Drug likeness score for the synthesized [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol and [2-(1-furan-2-yl) quinolin-4-yl] methanol derivatives (4a-c, 5a-c and 10a-f)

Compounds	miLog P ^a	TPSA ^b	n-Atoms	n-ON ^c	n-OHNH ^d	n-Violation	n-roth ^e	MW ^f
4a	4.34	52.34	23	4	0	0	3	303.32
4b	5.00	52.34	24	4	0	0	3	337.76
4c	4.46	52.34	24	4	0	0	3	321.34
5a	3.85	46.26	21	3	1	0	2	275.31
5b	4.50	46.26	22	3	1	0	2	309.75
5c	3.97	46.26	22	3	1	0	2	293.30
10a	3.20	46.26	18	3	1	0	2	259.69
10b	3.33	46.26	18	3	1	0	2	304.14
10c	2.66	46.26	18	3	1	0	2	243.24
10d	3.42	46.26	19	3	1	0	2	273.72
10e	3.55	46.26	19	3	1	0	2	318.17
10f	2.88	46.26	19	3	1	0	2	257.26
Ascorbic acid	-1.40	107.22	12	6	4	0	2	176.12
Streptomycin	-4.87	324.42	40	18	15	3	9	580.59
Fluconazole	-0.12	81.6	22	7	1	0	5	306.28

Antioxidant studies

The antioxidant potential of synthesized compounds (4a-c, 5a-c and 10a-f) was determined as an index of pharmacological usefulness. Two *in vitro* models were used to evaluate antioxidant properties; radical scavenging and hydrogen peroxide scavenging activity. The antioxidant properties were expressed in terms of percentage inhibition and 50 percent inhibitory concentration (IC₅₀) values (table 5 and 6).

DPPH radical scavenging assay

The antioxidant potential of compounds (4a-c, 5a-c and 10a-f) was determined by *in vitro* model systems (Blois 1958) and were used for the evaluation of antioxidant properties, namely, 2, 2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity and expressed in terms of 50% inhibitory concentration (IC₅₀) values (table 6).

Table 5: Antioxidant activity of substituted [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol and [2-(1-furan-2-yl) quinolin-4-yl] methanol derivatives (4a-c, 5a-c and 10a-f)

Compounds code	DPPH scavenging activity % inhibition	H ₂ O ₂ scavenging activity % inhibition
4a	68±0.57	64±1.0
4b	71±1.0	70±0.57
4c	82±1.52	68±0.57
5a	65±1.52	69±1.0
5b	74±1.0	70±2.5
5c	63±1.72	58±1.0
10a	56±1.52	52±3.02
10b	46±1.0	45±1.52
10c	85±1.52	72±1.72
10d	57±1.0	52±0.57
10e	61±1.15	64±1.52
10f	79±1.52	69±2.64
Ascorbic acid	96±1.15	76±1.0

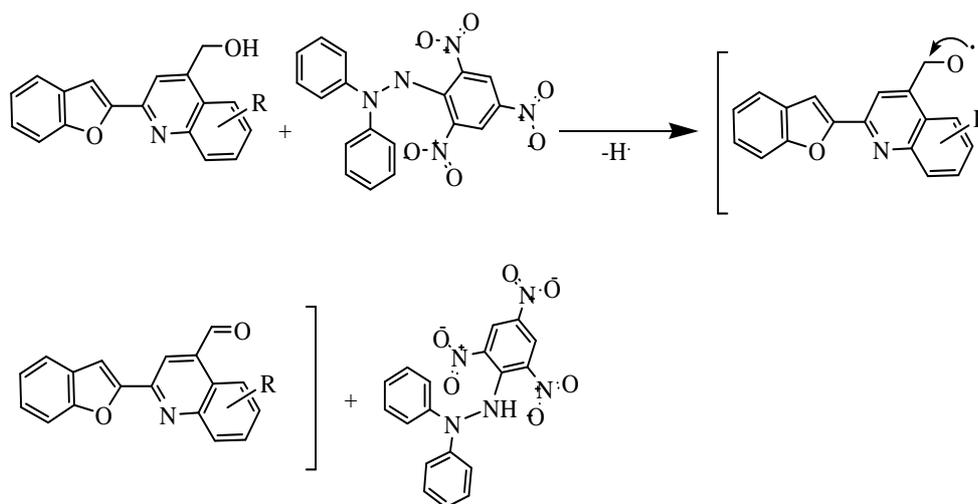
Values are expressed as mean±SD (n=3). Values are significant with each other at P<0.05 (Duncan's multiple range test).

The DPPH radical scavenging assay is one of the best free radicals scavenging mechanism (Scheme-III) by which antioxidant inhibits the oxidation and offer a rapid technique for screening the radical scavenging activity of specific compounds. All tested molecules exhibited a certain degree of radical scavenging activity. The compounds 4c, 5b, 10c and 10f exhibited good radical scavenging potential (table 5) compared with standard, ascorbic acid.

Hydrogen peroxide (H₂O₂) scavenging capacity assay

The hydrogen peroxide scavenging ability of synthesized derivatives was determined according to the method of Ruch and co-workers 1989 [42].

A solution of H₂O₂ (40 mM) was prepared in phosphate buffer (pH 7.4).



Scheme III: The mechanism for the reaction of compounds 5a-c with DPPH radical

Table 6: IC₅₀ (µg/ml) values of DPPH and H₂O₂ scavenging activity of substituted [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol and [2-(1-furan-2-yl) quinolin-4-yl] methanol analogues (4a-c, 5a-c and 10a-f)

Compounds code	DPPH scavenging activity IC ₅₀ (µg/ml)	H ₂ O ₂ scavenging activity IC ₅₀ (µg/ml)
4a	16.8±0.57	16±1.52
4b	17.4±0.25	15.5±0.20
4c	20±1.0	16±1.15
5a	16.5±0.2	15±1.0
5b	18±1.15	14.5±0.23
5c	16.5±0.2	9.8±0.11
10a	11.5±0.23	11±0.20
10b	9.5±0.20	8.2±1.15
10c	24±0.57	18±1.0
10d	12±1.52	11.5±0.20
10e	16±1.15	15±1.52
10f	13.8±0.15	13.2±0.30
Ascorbic acid	6±1.52	7.5±0.20

Values are expressed as mean±SD (n=3). Values are significant with each other at P<0.05 (Duncan's multiple range test).

The solutions of different molecules (10-50 µg/ml) were prepared in phosphate buffer and were added to H₂O₂ solution (0.6 ml, 40 mM). The absorbance value of the reaction mixture was recorded at 230 nm. The blank solution contains phosphate buffer without H₂O₂. The percentage of an H₂O₂ scavenging of the entitled molecule and the standard compound was calculated as H₂O₂ radical scavenging activity.

$$(\%) = \left[\frac{A_0 - A_1}{A_0} \right] \times 100$$

Where A₀ is the absorbance of H₂O₂, A₁ is the absorbance of H₂O₂ solution in the presence of benzofuran/furan quinolone methanols.

The compounds 4b, 5a, 10c and 10f exhibited good radical scavenging potential when compared with standard, ascorbic acid.

Antibacterial activity

The *in vitro* antibacterial activity of entitled compounds was determined by the agar well diffusion method. In this study, *E. coli*, *K. pneumonia* and *S. typhirium* (Gram-negative) were selected because of its infectious nature. The test compounds were dissolved in dimethyl sulfoxide (DMSO) at concentrations of 5 and 10 µg/ml (table 7).

Table 7: Antimicrobial activities of substituted [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol and [2-(1-furan-2-yl) quinolin-4-yl] methanol analogues (4a-c, 5a-c and 10a-f)

Samples code	<i>E. coli</i>		<i>K. pneumonia</i>		<i>S. typhimurium</i>	
	5 µg/ml	10 µg/ml	5 µg/ml	10 µg/ml	5 µg/ml	10 µg/ml
4a	2.4±0.14	2.9±0.11	3.2±0.08	3.4±0.14	2.5±0.11	3.0±0.11
4b	2.6±0.17	3.1±0.17	2.8±0.12	3.4±0.17	2.3±0.12	3.0±0.17
4c	3.1±0.12	3.5±0.21	2.7±0.17	3.6±0.26	3.4±0.12	3.6±0.31
5a	3.4±0.17	3.9±0.12	3.2±0.12	3.6±0.26	3.0±0.09	3.4±0.20
5b	3.3±0.17	3.6±0.18	2.8±0.09	3.5±0.22	3.1±0.2	3.6±0.12
5c	2.4±0.25	3.0±0.18	3.1±0.12	3.4±0.08	3.3±0.17	3.8±0.35
10a	2.6±0.17	3.4±0.26	3.0±0.12	3.5±0.12	2.8±0.10	3.3±0.17
10b	3.4±0.17	3.6±0.12	2.8±0.15	3.2±0.12	3.1±0.12	3.4±0.21
10c	2.4±0.27	2.8±0.18	3.1±0.12	3.3±0.18	3.0±0.12	3.5±0.17
10d	3.2±0.26	3.5±0.09	3.4±0.21	3.8±0.35	2.7±0.12	2.9±0.12
10e	2.7±0.31	3.1±0.12	3.2±0.20	3.6±0.12	3.4±0.22	3.8±0.40
10f	2.8±0.12	3.4±0.12	3.0±0.12	3.6±0.25	2.5±0.22	3.0±0.08
Streptomycin	2.7±0.12	3.8±0.09	3.4±0.21	3.6±0.26	3±0.12	3.5±0.18

Values are expressed as mean±SD (n=3). Values are significant with each other at P<0.05 (Duncan's multiple range test).

The results indicated that among the tested compounds, concerning antibacterial activities, compounds 4c, 5a-b, 10b 10d and 10f found to be potent against *E. coli*, while the compounds 4a-b, 5c, 10a, 10c and 10e shown significant activity. The compounds 4a, 5a and 10c-d found to be potent against *K. pneumonia*, while the compounds 4b-c, 5b-c, 10a-b and 10e-f shown moderate activity. The compounds 4c, 5a and 10c-d found to be possessed good inhibition against *S. typhimurium*, while the compounds 4a-b, 5b-c, 10a-b and 10e-f shown significant activity as compared with standard drug streptomycin. From the results, it could be accomplished that, the chloro and fluoro functioned derivatives showed better activity than un-substituted derivatives against all bacterial strains.

Antifungal activity

Further, the synthesized compounds were screened for their *in vitro* antifungal activity against *A. flavus* and *C. reoformans* because of their transmittable nature. The compounds were dissolved in DMSO and antifungal activity was determined by the poisoned food technique at concentrations of 50 and 100 µg/ml.

The antifungal result data (table 8) indicated that the synthesized compounds showed a variable degree of inhibition against fungal species, the compounds 4c, 5c and 10c-e showed potent activity against *A. flavus*, while the compounds 4a-b, 5a-b, 10a-b and 10f have shown moderate activity.

Table 8: Antifungal activities of substituted [2-(1-benzofuran-2-yl) quinolin-4-yl] methanol and [2-(1-furan-2-yl) quinolin-4-yl] methanol analogues (4a-c, 5a-c and 10a-f)

Samples code	<i>A. flavus</i> % inhibition		<i>C. neoformans</i> % inhibition	
	50 µg/ml	100 µg/ml	50 µg/ml	100 µg/ml
4a	36±1.52	60±1.0	54±1.52	86±2.08
4b	45±1.52	68±1.0	56±3.5	80±0.57
4c	56±1.0	79±0.57	36±1.15	85±1.0
5a	42±1.52	70±2.08	48±1.52	76±1.0
5b	47±1.0	78±2.0	54±1.15	87±1.15
5c	56±1.52	68±0.57	41±1.0	84±1.52
10a	34±1.0	65±1.52	54±2.0	90±1.52
10b	41±2.0	70±1.15	50±1.52	86±3.0
10c	51±3.5	68±2.0	36±1.15	71±2.08
10d	53±1.52	74±2.0	54±3.5	86±3.0
10e	57±1.0	78±1.52	68±0.57	80±2.0
10f	46±1.15	82±2.0	44±3.5	74±1.73
Fluconazole	47±0.57	64±2.3	42±0.57	75±0.57

Values are expressed as mean±SD (n=3). Values are significant with each other at P<0.05 (Duncan's multiple range test).

The compounds 4a-b, 5a-b, 10a-b and 10d showed good activity against *C. reoformans*, while the compounds 4c, 5c, 10c and 10f have showed significant activity when compared with standard drug fluconazole. The results exhibited that, the halogen-substituted derivatives showed better activity compared to other analogues.

Statistical analysis

The measurements were expressed as mean±SD for standard drugs. The data were analyzed using one-way analysis of variance (ANOVA) followed by Duncan's Multiple Range Test (DMRT) by using statistical package of social science (SPSS) version 10.0 for

Windows. A difference in the mean values of $P < 0.05$ was considered to be statistically significant.

CONCLUSION

The present work reports the synthesis, spectral characterization and biological activities including antioxidant and antimicrobial activities of synthesized series of benzofuran/furan quinoline methanol analogues (table 1). Molecules were made to predict ADMET/SAR *in silico*, the synthesized molecules are in an acceptable range were further screened for antioxidant analysis with DPPH and H_2O_2 scavenger methods and antimicrobial activity. The work indicates that the entitled compounds were found to possess good antioxidant and potent antimicrobial activity. The potent molecules (4b-c, 5a-c, and 10b-f) will be further taken up for in detail study, by varying the functionalities on the aromatic rings with a change in carbon chain length, the electron donor and withdrawing groups. The furan quinoline methanols can be considered as lead moieties for drug finding. The results provide useful information for operating as a positive reinforcement of the tendency to use antioxidant and antimicrobial properties as a guideline of the rational design of this class of compounds. It needs further detailed investigations such as *in vivo* pharmacokinetic profile, toxicity, the mechanism(s), are needed to evaluate their potential of developing into the therapeutic agents.

AUTHORS CONTRIBUTION

Dr. N. D. Satyanarayan is the mentor involved in designing and is the supervisor of the overall work. Mr. Anantacharya R is mainly involved in synthesis/characterization of entitled molecules and carried out *in silico* pharmacokinetic studies and their interpretation. Dr. K. M. Mahadevan involved in the idea generation of in the generation of new coupled heterocyclic analogues; Mr. Sameer P. was instrumental in carrying out *in vitro* antimicrobial and antioxidant results. Mr. Adarsha is helped in generating the spectral data.

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CONFLICT OF INTERESTS

There is no any conflict of interest

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